Antibacterial potential of synthesized iron oxide nanoparticles against uropathogens (*Escherichia coli* **and** *Staphylococcus aureus***)**

By

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Abstract

Background : The biosynthesis of metal nanoparticles is an important broad and developing field of research in many fields, especially medical fields. **Material and methods:** in current study, iron oxide nanoparticles (Fe₃O₄-NP_S) were synthesized from aqueous ferric chloride in a clear, simple, biosynthetic way using the aqueous extract of *Curcuma longa* powder, which is considered at the same time a reductant and a stabilizer as well. The prepared samples were characterized using several techniques: UV-Visible, TEM, SEM, and FT-IR. Through the color change, the formation of $Fe₃O₄$ -NPs is evident through the appearance of the characteristic black color of the solution. Formation of Fe from *C. Longa* was also determined by UV spectroscopy where surface plasmon absorption was observed at $223-303$ nm. Well-dispersed Fe₃O₄-NPs with isotropic and anisotropic morphology are seen for 20 mL of *C. longa* water extract with a size of 40 nm in the TEM images. The functional groups present in the NPs were mainly –OH and –COOH identified using FT-IR which makes them hydrophilic and hence the NPs do not need any functional modification. This study included bacterial isolates obtained from urine samples of patients suffering from urinary tract infections (UTIs), and then the antimicrobial activity of Fe₃O₄-NPs was evaluated against Gram-positive (*Staphylococcus aureus*) and Gram-negative (*Escherichia coli*) bacteria, and it was found that the zone of inhibition was 10.5 mm for (*E. coli*) and 17.6 mm for (*Staphylococcus aureus*). Thus naturally stabilized Fe₃O₄-NPs with herbal properties can be used in various biological applications.

Keywords: Green synthesis, *Curcuma longa*, Iron Oxide Nanoparticles (Fe₃O₄), urinary tract infections (UTIs)

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Introduction

A urinary tract infection (UTI) often happens when a pathogenic microbe attaches to and multiplies in a portion of the urinary system due to a breakdown in the host's defense mechanisms. There are many host defenses, including normal urination, mucosal barrier, anatomical structures as well as urine properties and systemic immune competence. Although fungi and viruses can also infect the urinary system, bacteria are the primary cause of UTIs [1]. Urinary tract infections can affect multiple anatomic sites. They fall into two categories: lower urinary tract infections, which affect the bladder, urethra, and vagina, and upper urinary tract infections, which affect the kidneys and ureters. The upward movement of germs through the urethra and reproductive system to the bladder, ureters, and one or both kidneys causes bacterial urinary tract infections [2,3].

Numerous types of bacterial strains become resistant to antibiotics as a result of the frequent and careless use of antibiotics, and thus treating the simplest types of bacterial infections have become very difficult. This is because novel bacterial strains that are extremely resistant to antibiotics are starting to appear. Therefore, scientists are always searching for new resistant and antibacterial sources that are more effective to overcome this phenomenon. Moreover, nanotechnology is considered a good alternative for this purpose [4,5,6,7].

We believe that the use of nanotechnology can significantly improve antibacterial therapy using iron oxide nanoparticles $(Fe₃O₄)$ to support cellular growth, and through food some antioxidants can be delivered exogenously and are referred to as exogenous antioxidants. Recently, there has been a lot of interest in the creation of biocompatible nanocomposites with antioxidant capabilities and prospective applications in the medical domain [8,9,10,11]. Iron oxide nanoparticles, which range in size from 1 to 100 nm, are a relatively new invention that is gaining a lot of attention and showing promise for use in numerous medical application studies [12,13,14,15]. Characteristics of magnetism, including strong coercivity, high magnetic susceptibility, and supermagnetism [16]. The primary objective of this research is to investigate the antibacterial activities of chemically produced iron oxide nanoparticles (Fe3O4) against bacterial strains such as *Escherichia coli* and *Staphylococcus aureus*.

Materials and methods

Materials

The phosphate buffer solution (PBS) and iron (II) chloride (FeCl₂• $4H_2O$), which were acquired from Sigma-Aldrich Company in India, are the chemicals utilized in this experiment. The suppliers of the culture medium included Himedia Company in India for Gram stain and blood agar, and Accumedia Company in the USA for MacConkey agar, Mannitol salt agar, and nutritional broth.

Isolation and Identification of Microorganisms

From hospitalized UTI patients' urine samples have been collected and cultivated on selective culture media, the disease-causing bacteria (*Escherichia coli* and *Staphylococcus aureus*) were identified based on their phenotypic traits through several biochemical tests and by using the Vitek2 system for the final diagnosis of the isolates. The identification of bacterial morphology was

determined using gram stain and colony morphology after cultivation on the following culture media MacConkey agar and Mannitol salt agar [17,18].

Extraction of *Curcuma longa* **Solution**

In a 200 ml beaker, 10 g of *C. longa* powder was dissolved in 100 ml of deionized (D.I.) water, and the mixture was agitated for four hours at 70 °C [19]**.** The *C. longa* solution was then extracted as shown in Figure 1.

Figure 1: *Curcuma longa* extract. **Synthesis of (Fe3O4-NPs) from (FeCl2.4H2O).**

Firstly, $Fe₃O₄$ -NPs were prepared, the synthesis part was carried in the laboratories of the University of Baghdad. First, 2.7 g of Iron (II) Chloride (FeCl₂· 4H₂O) was dissolved in 100 mL of deionized water (D.I.). Then 50 ml of *C. longa* solution was added with continuous stirring. The resulting solution was heated at 80 °C; the stirring was continued for about three hours as shown in Figure 2. After the reaction was completed, the solution was placed in a centrifuge to obtain a precipitate. The precipitate was washed several times with D.I. and then the precipitated black nanoparticles were dried [20]. Secondly, tests were conducted to characterize the composition of the nanocomposite, in order to determine the nanoparticles' dimensions and formation, as well as to study the morphological characteristics of the iron oxide nanoparticles (Fe₃O₄).

Figure 2: Synthesis of (Fe₃O₄ - NPs)

http://xisdxjxsu.asia VOLUME 20 ISSUE 08 AUGUST 2024**836-846 Measurement of Fe3O4-NPs' Antibacterial Activity**

The well diffusion method was used to detect the biological activity of the prepared nanoparticles. Following the manufacturer's instructions. Iron oxide nanoparticles (Fe₃O₄) were used at concentration 100 μ g mL⁻¹, at a temperature of 37 °C and incubated for 24 hours. The resulting diameter of inhibition was then recorded [21,22] Reference drug – Gentamicin 100 μ g mL⁻¹.

Results and Discussion

Synthesis and Characterization of Fe3O4-NPs

The outcomes demonstrated the successful synthesis of iron oxide nanoparticles (Fe₃O₄), which possessed ferromagnetic characteristics and a dark black hue as depicted in Figure (3). The obtained results were consistent with those of a previous study conducted by Win et al. [23].

Figure 3: Characterization of (Fe₃O₄ - NP_S)

Characterization of (Fe3O4-NPs)

The Precise scanning electron microscope (SEM) analysis is widely used to identify the surface morphology of the bio-synthesized nanoparticles. SEM images showed the production of iron oxide nanoparticles (Fe₃O₄) as presented in Figure 4. It can be observed from the SEM images that FeO nanoparticles had spherical morphology with little aggregation, and the particles were within the size range of 30 to 55 nm. Since the dimensions of the nanoparticles greatly influence the properties of the nanocrystals, and controlling the monodispersed size is also essential [24]. Morphology and size characterization of Fe₃O₄-NPs were also examined by microscopic analysis using transmission electron microscopy (TEM). Diameter measurements with TEM were performed using Image J software as depicted in Figure 5. TEM images showed that iron oxide nanoparticles were spherical and had an average size of 40 nm. Similar findings were reported by Mendes et al. [25] who have synthesized spherical shaped FeO NPs with different sizes.

Figure 4: SEM images of iron oxide nanoparticles (Fe₃O₄) size and top view.

Figure 5: TEM image of (Fe₃O₄-NPs)

FT-IR Analysis of (Fe3O4-NPs)

FT-IR analysis was performed to identify the biomolecules responsible for the reduction of iron oxide nanoparticles present in *Curcuma longa* extract. Based on the current results, FT-IR analysis gave clear stretching vibrations at 3400.5 cm⁻¹,1625.99 cm⁻¹ and 626.87 cm⁻¹ in the region of 400 -4000 cm⁻¹ (Fig. 6). These peaks represent the following bonding in the sample confirms the reducing agent role in the formation of Fe₃O₄ –NPs. The broad peak around 3400.5 cm⁻¹ showed the OH stretching bond vibration which was due to the water adsorption on the surface of $Fe₃O₄ - NPs$ [26], while 1625.99 cm^{-1} corresponds to the stretching of the C=O bond to the phytochemicals present in the plant extract. The present findings seem to be consistent with a previous study which found that the presence of bioactive compounds may be responsible for the capping, reduction and conversion of Fe⁺ to Fe-O nanoparticles [27]. In addition, a broad short peak appears at 626.87 cm⁻¹

corresponds to the inorganic stretching of Fe–O–Fe and indicates the formation of iron oxide nanoparticles [28].

Figure 6: FT-IR analysis of (Fe₃O₄-NPs)

UV-Visible Spectrophotometer of (Fe3O4-NPs)

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UV-Visible spectrum of iron oxide nanoparticles is shown in Figure 7. As is widely known, metal oxide NPs have significant absorption bands between 200 and 800 nm at room temperature. In the present study, UV-visible spectra of the synthesized $Fe₃O₄$ -NPs showed a characteristic absorption peak between 223 and 303 nm. It was reported earlier that the UVspectral absorbance around 250-260 nm is a unique characteristic feature of Fe₃O₄-NPs [29]. According to the obtained data, the absorption maxima of synthesized iron oxide nanoparticles are similar to those observed in other studies which used *Ficus carica* and *Skimmia laureola* extracts to produce iron nanoparticles [30-31].

Figure 7: UV-visible spectrophotometer of (Fe₃O₄-NPs)

Antimicrobial Activity of (Fe3O4-NPs) Against Microbial Isolates

http://xisdxjxsu.asia VOLUME 20 ISSUE 08 AUGUST 2024**836-846** The agar well-diffusion method was used to evaluate the antibacterial activity of $Fe₃O₄$ -NPs at a concentration of 100 µg mL−1 against Gram-negative (*Escherichia coli*) and Gram-positive (*Staphylococcus aureus*) bacteria as demonstrated in Figure 8. According to the obtained data, the antibacterial property of the bio-synthesized $Fe₃O₄-NPs$ revealed that the nanoparticles have moderate antibacterial activity in comparison to the used antibiotic. The result also showed higher antibacterial activity against *S. aureus*, whereas moderate activity was revealed against *E. coli*. The inhibitory activities in culture media of the $Fe₃O₄$ -NPs reported in Table. 1. Although, the

antibacterial activities of several bio-synthesized iron oxide nanoparticles have been reported, the bacteria inhibition mechanism of the nanoparticles has yet to be elucidated. However, some theories have suggested that the nanoparticles invade the cell membrane and denature the enzyme of bacteria, and damage bacterial DNA, which in turn induce cell death (32,33). It has been reported that bio-synthesized nanoparticles show better antimicrobial activity compared to chemically synthesized nanoparticles, because various plant extracts that were used for the synthesis of nanoparticles have antimicrobial properties [23].

Table 1: Antimicrobial Activity of the Synthesized Iron Oxide Nanoparticles by *Curcuma longa* Aqueous Extract.

Concluding Remarks

The most common microbes isolated from samples of patients with urinary tract infections were (*Escherichia coli*) and (*Staphylococcus aureus*). Since physiological activities occur at the nanoscale, the attachment of iron oxide $(Fe₃O₄)$ nanoparticles is expected to alleviate many biological and healthcare difficulties. It's known that the uncontrolled use of antibiotics might fuel the growth of resistant strains around the world. The synthesis of NPs is considered a powerful

technology due to their potency against common multidrugresistant (MDR) species. Our findings demonstrated the successful synthesis of iron oxide (Fe₃O₄) nanoparticles and their application for antibacterial activity against bacterial isolates that are resistant to multiple

drugs and antioxidants. We may conclude that superparamagnetic and very potent antibacterial agents are present in iron oxide nanoparticles $(F_{20}Q_4)$.

Conflict of Interest

The research's publishing is free of conflicts of interest, according to the authors.

References

- 1) Wagenlehner FM, Bjerklund Johansen TE, Cai T, Koves B, Kranz J, Pilatz A, Tandogdu Z. Epidemiology, Definition and Treatment of Complicated Urinary Tract Infections. Nature Reviews Urology. 2020; 17(10):586-600. <https://doi.org/10.1038/s41585-020-0362-4>
- 2) Storme, O., Tirán Saucedo, J., Garcia-Mora, A., Dehesa-Dávila, M. and Naber, K.G. Risk Factors and Predisposing Conditions for Urinary Tract Infection. Therapeutic Advances in Urology. 2019; 11:10-28. <https://doi.org/10.1177/1756287218814382>
- 3) Hickling, D.R., Sun, T.T. and Wu, X.R. Anatomy and Physiology of the Urinary Tract: Relation to Host Defense and Microbial Infection. Urinary Tract Infections: Molecular Pathogenesis and Clinical Management. American Society for Microbiology, pp.1-25. <https://doi.org/10.1128/9781555817404.ch1>
- 4) Terreni, M., Taccani, M. and Pregnolato, M. New Antibiotics for Multidrug-Resistant Bacterial Strains: Latest Research Developments and Future Perspectives. Molecules. 2021; 26(9):1-31. <https://doi.org/10.3390/molecules26092671>
- 5) Mancuso, G., Midiri, A., Gerace, E. and Biondo, C. Bacterial Antibiotic Resistance: The Most Critical Pathogens. Pathogens. 2021; (10):1-14. <https://doi.org/10.3390/pathogens10101310>
- 6) Gómez-Núñez, M.F., Castillo-López, M., Sevilla-Castillo, F., Roque-Reyes, O.J., Romero-Lechuga, F., Medina-Santos, D.I., Martínez-Daniel, R. and Peón, A.N. Nanoparticle-Based Devices in the Control of Antibiotic Resistant Bacteria. Frontiers in Microbiology. 2020; (11):1-15. <https://doi.org/10.3389/fmicb.2020.563821>
- 7) Kadhum MM, Hussein NN. Detection of the Antimicrobial Activity of Silver Nanoparticles Biosynthesized by *Streptococcus pyogenes* Bacteria. Iraqi Journal of Agricultural Sciences. 2020; 51(2):500-507. <https://www.iasj.net/iasj/download/5deda042c2d368df>
- 8) Xu C, Akakuru OU, Zheng J, Wu A. Applications of Iron Oxide-Based Magnetic Nanoparticles in the Diagnosis and Treatment of Bacterial Infections. Frontiers in Bioengineering and Biotechnology. 2019; (7):1-15. <https://doi.org/10.3389/fbioe.2019.00141>
- 9) Abu-Huwaij R, Al-Assaf SF, Mousli F, Kutkut MS, Al-Bashtawi A. Perceptive Review on Properties of Iron Oxide Nanoparticles and their Antimicrobial and Anticancer Activity. Sys. Rev. Pharm. 2020; 11(8):418-31. <https://doi.org/10.1111/jocd.14704>
- 10) Rukhsar M, Ahmad Z, Rauf A, Zeb H, Ur-Rehman M, Hemeg HA. An Overview of Iron Oxide (Fe3O4) Nanoparticles: From Synthetic Strategies, Characterization to Antibacterial and Anticancer Applications. Crystals. 2022; 12(12):1809. <https://doi.org/10.3390/cryst12121809>
- 11) Abdulsada FM, Hussein NN, Sulaiman GM. Potentials of Iron Oxide Nanoparticles (Fe3O4): As Antioxidant and Alternative Therapeutic Agent Against Common Multidrug-Resistant Microbial Species. Iraqi Journal of Science. 2023;64(6):2759-73. [DOI: 10.24996/ijs.2023.64.6.10](oo)
- 12) Silva AK, Espinosa A, Kolosnjaj‐Tabi J, Wilhelm C, Gazeau F. Medical Applications of Iron Oxide Nanoparticles. Iron Oxides: From Nature to Applications. 2016; (18):425-72. <https://doi.org/10.1002/9783527691395.ch18>
- 13) Natarajan S, Harini K, Gajula GP, Sarmento B, Neves-Petersen MT, Thiagarajan V. Multifunctional Magnetic Iron Oxide Nanoparticles: Diverse Synthetic Approaches, Surface Modifications, Cytotoxicity Towards Biomedical and Industrial Applications. BMC Materials. 2019;1(1):1-22. <https://doi.org/10.1186/s42833-019-0002-6>
- 14) Sahu T, Ratre YK, Chauhan S, Bhaskar LV, Nair MP, Verma HK. Nanotechnology Based Drug Delivery System: Current Strategies and Emerging Therapeutic Potential for Medical Science. Journal of Drug Delivery Science and Technology. 2021; (63):102487. <https://doi.org/10.1016/j.jddst.2021.102487>
- 15) Li Y, Ye D, Li M, Ma M, Gu N. Adaptive Materials Based on Iron Oxide Nanoparticles for Bone Regeneration. ChemPhysChem. 2018;19(16):1965-79. <https://doi.org/10.1002/cphc.201701294>
- 16) Upadhyay S, Parekh K, Pandey B. Influence of Crystallite Size on the Magnetic Properties of Fe3O⁴ Nanoparticles. Journal of Alloys and Compounds. 2016; 678:478-485. <https://doi.org/10.1016/j.jallcom.2016.03.279>
- 17) Ghaib NH, Al-Ghurabi BH, Hassan SS. Isolation of Moraxella and Streptococcus from New Packed Orthodontic Materials. Journal of Global Pharma Technology. 2019; 11(3): 533-535. <www.jgpt.co.in>
- 18) Hassan SS, Ghaib NH, Al-Ghurabi BH. Assessment of Bacterial Contamination of Orthodontic Arch Wire. Journal of Baghdad College of Dentistry. 2019;31(1):48-51. <https://doi.org/10.26477/jbcd.v31i1.2578>
- 19) Momenkiaei F, Raofie F. Preparation of *Curcuma longa L*. Extract Nanoparticles Using Supercritical Solution Expansion. Journal of Pharmaceutical Sciences. 2019;108(4):1-9. <https://doi.org/10.1016/j.xphs.2018.11.010>
- 20) Nguyen MD, Tran HV, Xu S, Lee TR. Fe3O⁴ Nanoparticles: Structures, Synthesis, Magnetic Properties, Surface Functionalization, and Emerging Applications. Applied Sciences. 2021;11(23):11301. <https://doi.org/10.3390/app112311301>
- 21) Pourebrahim F, Ghaedi M, Dashtian K, Kheirandish S, Jannesar R, Pezeshkpour V. Preparation of Chitosan Functionalized End‐Capped Ag‐NPs and Composited with Fe3O4‐NPs: Controlled Release

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to pH‐Responsive Delivery of Progesterone and Antibacterial Activity Against *Pseudomonas aeruginosa* (PAO‐1). Applied Organometallic Chemistry. 2018;32(1):1-10. <https://doi.org/10.1002/aoc.3921>

- 22) Ansari SA, Oves M, Satar R, Khan A, Ahmad SI, Jafri MA, Zaidi SK, Alqahtani MH. Antibacterial Activity of Iron Oxide Nanoparticles Synthesized by Co-Precipitation Technology Against *Bacillus cereus* and *Klebsiella pneumoniae*. Polish Journal of Chemical Technology. 2017;19(4):110-115. <http://doi.org/10.1515/pjct-2017-0076>
- 23) Win TT, Khan S, Bo B, Zada S, Fu P. Green Synthesis and Characterization of $Fe₃O₄$ Nanoparticles Using Chlorella-K01 Extract for Potential Enhancement of Plant Growth Stimulating and Antifungal Activity. Scientific Reports. 2021;11(1):21996. <https://doi.org/10.1038/s41598-021-01538-2>
- 24) Mahdavi M, Namvar F, Ahmad MB, Mohamad R. Green Biosynthesis and Characterization of Magnetic Iron Oxide (Fe3O4) Nanoparticles Using Seaweed (*Sargassum muticum*) Aqueous Extract. Molecules. 2013;18(5):5954-64. <https://doi.org/10.3390/molecules18055954>
- 25) Mendes RG, Koch B, Bachmatiuk A, El-Gendy AA, Krupskaya Y, Springer A, Klingeler R, Schmidt O, Büchner B, Sanchez S, Rümmeli MH. Synthesis and Toxicity Characterization of Carbon Coated Iron Oxide Nanoparticles with Highly Defined Size Distributions. Biochimica et Biophysica Acta (BBA)-General Subjects. 2014;1840(1):160-169. <https://doi.org/10.1016/j.bbagen.2013.08.025>
- 26)Jamzad M, Kamari Bidkorpeh M. Green Synthesis of Iron Oxide Nanoparticles by the Aqueous Extract of *Laurus nobilis* L*.* Leaves and Evaluation of the Antimicrobial Activity. Journal of Nanostructure in Chemistry. 2020;10(3):193-201. <https://doi.org/10.1007/s40097-020-00341-1>
- 27) Bibi I, Nazar N, Ata S, Sultan M, Ali A, Abbas A, Jilani K, Kamal S, Sarim FM, Khan MI, Jalal F. Green Synthesis of Iron Oxide Nanoparticles Using Pomegranate Seeds Extract and Photocatalytic Activity Evaluation for the Degradation of Textile Dye. Journal of Materials Research and Technology. 2019;8(6):6115-24. <https://doi.org/10.1016/j.jmrt.2019.10.006>
- 28) Herrera-Becerra R, Zorrilla C, Rius JL, Ascencio JA. Electron Microscopy Characterization of Biosynthesized Iron Oxide Nanoparticles. Applied Physics A. 2008; 91:241-6. [https//doi.org/10.1007/s00339-008-4420-7](Documents)
- 29) Mirza AU, Kareem A, Nami SA, Khan MS, Rehman S, Bhat SA, Mohammad A, Nishat N. Biogenic Synthesis of Iron Oxide Nanoparticles Using *Agrewia optiva* and *Prunus persica* Phyto Species: Characterization, Antibacterial and Antioxidant Activity. Journal of Photochemistry and Photobiology B: Biology. 2018; 185:262-74. <https://doi.org/10.1016/j.jphotobiol.2018.06.009>
- 30) Tahir A, Saeed A, Ramzan I, Hayat SS, Ahmad W, Naeem S, Afzal M, Mukhtar A, Mehmood T, Khan BS. Mechanism for the Formation of Magnetite Iron Oxide Nanostructures by *Ficus carica* Dried Fruit Extract Using Green Synthesis Method. Applied Nanoscience. 2021;11(6):1857-65. [https//doi.org/10.1007/s13204-021-01860-1](Documents)

31) Alam T, Khan RA, Ali A, Sher H, Ullah Z, Ali M. Biogenic Synthesis of Iron Oxide Nanoparticles Via *Skimmia laureola* and their Antibacterial Efficacy against Bacterial Wilt Pathogen *Ralstonia solanacearum*. Materials Science and Engineering: C. 2019; 98:101-8. <https://doi.org/10.1016/j.msec.2018.12.117>