

A Review of Different Medicinal Plants with Potential Anti-inflammatory Activities

Muhammad Naseer, Muhammad Adil, Faiza Tawab², Ayaz Ali Sher¹, Ambrin¹, Fakhre Alam¹

Department of Chemical and Life Sciences, Qurtuba University of Science and Information Technology, Peshawar, Pakistan

Department of Botany, Shaheed Benazir Bhutto Women University, Peshawar

Abstract

Numerous nonsteroidal anti-inflammatory drugs have been shown to alleviate pain and inflammation by inhibiting the cyclooxygenase enzyme isoform, thereby reducing the synthesis of prostaglandins from arachidonic acid. Unfortunately, the use of these drugs is associated with a range of adverse effects. However, there exists a group of medicinal plants that exhibit anti-inflammatory therapeutic properties with minimal or no adverse effects. These plants have proven successful in treating inflammatory disorders in conventional medicine. It's noteworthy that researchers have extensively studied and documented the biological and therapeutic effects of some common medicinal plants. Regrettably, there is a notable absence of a comprehensive review paper cataloging medicinal plants with anti-inflammatory properties across various African nations. Recognizing the significance of ethnobotanical knowledge, it is crucial to document and understand the applications of anti-inflammatory medicinal plants from selected countries representing different regions of the African continent. This approach aims to illustrate the diverse array of medicinal plants utilized for traditional or therapeutic purposes globally, and the present study enumerates the anti-inflammatory properties of several such plants from varied geographic regions

Keywords: Inflammation, Anti-inflammatory conditions, local plants, plant potentials.

Introduction

In various diseases, inflammation and oxidative stress have long been considered the primary pathological processes. Numerous studies have provided scientific evidence revealing the interconnected mechanisms governing inflammation induced by oxidative stress and vice versa. Inflammatory cells release reactive oxygen species (ROS), causing heightened oxidative damage at the inflammation site. Moreover, ROS and oxidative stress products amplify pro-inflammatory reactions. Researchers have increasingly turned to traditional knowledge of medicinal plants and their active components for developing new medications. Medicinal plants, due to their low toxicity, have been prioritized for testing and developing treatments for various ailments. Natural sources of antioxidants and anti-inflammatory compounds, such as polyphenol extracts, have gained attention (Liu *et al.*, 2022).

Over the years, medicinal plants have played a crucial role in the traditional treatment of numerous human diseases, including leprosy, gonorrhoea, dyspepsia, wounds, malaria, fever, and diarrhoea (Hussain *et al.*, 2021). The therapeutic effects of these plants are attributed to various phytochemicals, such as alkaloids, tannins, terpenoids, flavonoid glycosides, and saponins, which exhibit diverse physiological effects on humans (Junejo *et al.*, 2020). Utilizing plant-derived anti-inflammatory medications has been advocated as a logical and effective approach, as these compounds are known to suppress specific inflammatory mediators (Lahare *et al.*, 2021).

In response to harmful substances, living mammalian tissues often undergo local inflammation, a process that eliminates allergens or microorganisms and accelerates tissue restoration (Zhao *et al.*, 2021). Inflammation manifests as redness, swelling, heat, pain, and functional impairment. Although inflammation is a protective response, it is associated with various disease conditions at the cellular level, including arthritis, prostatitis, anaphylaxis, diabetes, chronic kidney disease, digestive disorders, Alzheimer's disease, bacterial infections, and novel coronavirus infections, leading to serious health issues (Bandyopadhyay *et al.*, 2021). Furthermore, it has been reported that inflammatory bowel disease (IBD), a chronic inflammatory disorder, affects over 3 million people in the United States, with 1.6 million residing in the region (Kumar, 2021).

Methods:

The research encompassed an extensive exploration of databases, including PubMed, Google Scholar, Scopus, and ScienceDirect. The search focused on identifying relevant studies incorporating terms related to inflammation, inflammatory conditions, therapeutic plants, and the potential of plants. The search duration was not restricted to ensure comprehensive coverage of pertinent materials. Information on isolated chemical compounds, pharmacological activity, utilized plant parts, experimental models, and other relevant details was systematically gathered and compiled.

Table 1: The medicinal plants which are considered to possess anti-inflammatory activity based on in vitro studies:

S. no	Scientific name	Family	Local name	Part/solvent used	Types of assays	Anti-inflammatory Activity %	IC50	Active compounds	Ref
1	<i>Aglaea borneensis</i>	Connaraceae	Akar rusa-rusa	Bark/methanol	LOX inhibition	71%–100% at 100 µg/m	NA	NA	Chung <i>et al.</i> , 2009
2	<i>Timonius flavescens</i>	Rubiaceae	Batut	Leaves/methanol	LOX inhibition	71%–100% at 100 µg/m	8.9 µg/m	NA	Chung <i>et al.</i> , 2009
3	<i>Anacardi um occidentale</i>	Anacardiaceae	Pokok gajus	Leaves/methanol	NO inhibition	16.10% at 250 µg/ml	NA	NA	Abas <i>et al.</i> , 2006
4	<i>Thymus vulgaris</i>	Lamiaceae	Taim	Whole plant/methanol	LOX inhibition	62% at 100 µg/ml	NA	NA	Abu Bakar <i>etal</i> 2018
5	<i>Averrhoa bilimbi</i>	Oxalidaceae	Belimbing buluh	Fruits/water	NO inhibition	22.30% at 250 µg/ml	NA	NA	Abas <i>et al.</i> , 2006

6	<i>Solanum torvum</i>	Solanaceae	Terung belanda	Leaves and fruits/methanol	NO inhibition	25.20% at 250 µg/ml	NA	NA	Abas <i>et al</i> 2006
7	<i>Barringtonia racemosa</i>	Lecythidaceae	Put at kampung	Leaves/ethanol	Griess assay (NO inhibition)	29.80% at 100 µg/ml	NA	NA	Ana., <i>et al.</i> , 2007
8	<i>Solanum nigrum</i>	Solanaceae	Terung meranti	Leaves/methanol	NO inhibition	27.60% at 250 µg/ml	NA	NA	Abas <i>et al.</i> , 2006

Table .1 Continue

s. no	Scientific name	Family name	Local name	Part/solvent used	Types of assays	Anti-inflammatory activity %	IC50	Active compounds	References
9	<i>Ficus deltoidea</i>	Moraceae	Mas cotek	Leaves /methanol	LOX inhibition	10.35% at 100 µg/ml	NA	NA	Abdullah <i>et al.</i> , 2009
10	<i>Eurycoma alinifolia</i>	Simaroubaeaceae	Tongkat ali	Root/ hydroalcohols	Human red blood cell	70.97% at 1000 µg/m	NA	NA	Varghese <i>et al.</i> , 2013
11	<i>Garcinia subelliptica</i>	Guttiferae	Pokok penanti	Seeds/ chloroform	Chemical mediator	NA	18.2 µM	Garsubellin	Weng., <i>et al.</i> , 2003
12	<i>Gynura pseudochina</i>	Asteraceae	Pokok daun dewa	Leaves/ ethyl acetate	IL-6/luciferase assay	NA	11.63 µg/m	NA	Siriwata nametan on, <i>et al.</i> , 2010
13	<i>Cyathostem aexcelsia</i>	Annonaceae	Liana	Leaves and stems/methanol	LOX inhibition	41%–70% at 100 µg/ml	NA	NA	Chungetal., 2009

14	<i>Jatropha acurcas</i>	Euphorbiaceae	Jarak pagar	Leaves/aqueous methanol	NO inhibition	NA	93.5 µg/ml	NA	Ehsan <i>et al.</i> , 2011
15	<i>Kaempferia agalanga</i>	Zingiberaceae	Cekur	petroleum ether, chloroform	COX-2 inhibitory screenin g assay	57.82% at 200 µg/ml	0.83 µM	NA	Umar <i>et al.</i> , 2012
16	<i>Musa acuminata</i>	Musaceae	Pisang abu nipah	Flowering stalk/methanol	(NO inhibition)	71.06% at 100 µg/ml	42.24 µg/ml	NA	Lee <i>et al.</i> , 2011
17	<i>Ocimum canum</i>	Lamiaceae	Kemangi putih	Whole plant/methanol	LOX inhibition	32% at 100 µg/ml	NA	NA	Abu Bakar <i>et al.</i> 2018

Table. 1 continue

s.no	Scientific name	Family	Local name	Part/solvent used	Types of assays	Anti-inflammatory activity %	IC50	A.compounds	Ref
18	<i>Piper sarmentosum</i>	Piperaceae	Kaduk	Leaves/methanol	Griess assay	62.82% at 100 µg/ml	60.24 µg/ml	NA	Lee <i>et al.</i> , 2011
19	<i>Curcuma longa</i>	Zingiberaceae	Kunyit	Rhizomes/hexane-ethyl	COX-2 inhibitory activity	58.90% at 125 µg/ml	NA	NA	Ram Sewake <i>et al.</i> , 2000.

20	<i>Boswellia serrata</i>	Burseraceae	Kemenyan	Leaves/methanol	Human red blood cell	80.00% at 2000 µg/ml	NA	NA	Afsar <i>et al.</i> , 2012
21	<i>Pithecellobium confertum</i>	Fabaceae	Medang	Seeds/methanol	NO inhibition	23.50% at 250 µg/m	NA	NA	Abas <i>et al.</i> , 2006
22	<i>Crinum asiaticum</i>	Amaryllidaceae	Pokok bakung	Leaves/ethanol	NO inhibition	NA	58.5 µg/ml	NA	Kim, <i>et al.</i> , 2008
23	<i>Chisocheton polyandrous</i>	Meliaceae	Lisi-lisi	Leaves/hexane	Soybean	NA	1.11 µM	Hydroxydammara	Chan., <i>et al.</i> , 2012
24	<i>Litsea garciae</i>	Lauraceae	Engkala/	Fruits/methanol	LOX assay	9.42% at 2 mg/ml	NA	NA	Kutoi <i>et al.</i> , 2012
25	<i>Carica papaya</i>	Caricaceae	Betik	Leaves/methanol	(NO inhibition)	72.63% at 100 µg/ml	60.18 µg/ml	NA	Lee <i>et al.</i> , 2011

Table. 1 continue

S.no	Scientific name	Family	Local name	Part/solvent used	Types of assays	Anti-inflammatory activity %	IC 50	Active compounds	Ref
26	<i>Leucas linifolia</i>	Lamiaceae	Ketumbak	Whole plant/methanol	LOX inhibition	34% at 100 µg/m	NA	NA	Abu Bakar <i>et al.</i> 2018
27	<i>Cosmos caudatus</i>	Asteraceae	Ulam raj	Leaves/methanol	NO inhibition	15.40% at 250 µg/ml	NA	NA	Abas. <i>et al.</i> , 2006
28	<i>Melicope ptelefolia</i>	Rutaceae	Tenggek burung	Leaves/methanol	NO inhibition	95% at 250 µg/ml	NA	p-O-geranylcoumaric	Shaari <i>et al.</i> , 2011

29	<i>Ficus deltoidei</i>	Moraceae	Mas cotek	Leaves/methanol	LOX inhibition	10.35 % at 100 µg/ml	NA	NA	Abdullah <i>et al.</i> , 2009
30	<i>Citrullus lanatus</i>	Cucurbitaceae	Tembika	chloroform, and 90% ethanol	COX-2 inhibitory activity	60–70% at 100 µM	69 µM	Cucurbitacin E	Abdel Wahab <i>et al.</i> , 2011
31	<i>Buchanania insignis</i>	Anacardiaceae	Tais/man gga hut an	Bark/methanol	LOX inhibition	41%–70% at 100 µg/ml	NA	NA	Chunget <i>et al.</i> , 2009
32	<i>Buchanania insignis</i>	Anacardiaceae	Tais/	Bark/methanol	LOX inhibition	41%–70% at 100 µg/ml	NA	NA	Chunget <i>et al.</i> , 2009
33	<i>Desmos chinensis</i>	Annonaceae	Kenanga hutan	Bark/methanol	LOX inhibition	41%–70% at 100 µg/ml	NA	NA	Chunget <i>et al.</i> , 2009

Table. 1 Continue

S no	Scientific name	Family	Local name	Part/solvent used	Types of assays	Anti-inflammatory activity %	IC50	Active compounds	Ref
34	<i>Canarium patentinervium</i>	Burseraceae	Kedondong	Leaves	5-LOX inhibition	NA	1.76 µM	Scopoletin	Mogana <i>et al.</i> , 2013
35	<i>Eurycoma longifolia</i>	Simaroubaceae	Tongkat ali	Root/hydroalcoholic	Human red blood cell	70.97 % at 1000 µg/ml	NA	NA	Varghese <i>et al.</i> , 2013
36	<i>Cyathostema excelsia</i>	Annonaceae	Lianas	Leaves	LOX inhibition	41%–70% at 100 µg/ml	NA	NA	Chunget <i>et al.</i> , 2009

37	<i>Boesenbergia rotunda</i>	Zingiberaceae	Temu kunci	Rhizomes/hexane	Griess assay	NA	36.68 μ M	Boesenbergin	Isa <i>et al.</i> , 2012
38	<i>Gynura pseudochina</i>	Asteraceae	Pokok daun dewa	Leaves/ethyl acetate	IL-6/luciferase assay	NA	11.63 μ g/ml	NA	Siriwa tanam et al., 2010
39	<i>Piper sarmentosum</i>	Piperaceae	Kaduk	Leaves/methanol	Griess assay	62.82% at 100 μ g/ml	60.24 μ g/ml	NA	Lee <i>et al.</i> , 2011
40	<i>Labiatae pumila var. alata</i>	Myrsinaceae	Kacip Fatimah	Roots/methanol	Colorimetric nitric oxide	75.68% at 100 μ g/ml	NA	NA	Karim <i>et al.</i> , 2013

Table 2: The medicinal plants which are considered to possess anti-inflammatory activity based on *in vivo* studies.

S no	Scientific name	Family	Local name	Part/solvent used	Dose of the extract	Experimental animals	Results	Ref
1	<i>Polygonum minus</i>	Polygonaceae	Kesum	Aerial parts/water	100 mg/kg and 300 mg/kg	Wistar albino rats	+e extracts significantly reduced the paw edema volume in the rats after 4 h	George <i>et al.</i> , 2014
2	<i>Annona muricata</i>	Annonaceae	Durian belanda	Leaves/aqueous ethanol	10–300 mg/kg	Sprague-Dawley rats	A significant decrease of the concentration of the proinflammatory cytokines TNF- α and IL-1 β was observed	Fong <i>et al.</i> , 2014

3	<i>Achyranthes aspera</i>	Amaranthaceae	Ara songsang	Root/ethyl alcohol	50, 100, and 200 mg/kg	Wistar rats	All the doses caused significant reduction in paw edema compared to control	Vijaya Kuma <i>ret al.</i> , 2009
4	<i>Ficus deltoidea</i>	Moraceae	Mas cotek	Whole plant/water	30, 100, and 300 mg/kg	Sprague-Dawley rats	The rats' paw edema volume reduced significantly in a dose-dependent manner	Zakaria <i>et al.</i> , 2012
5	<i>Sandoricum koetjape</i>	Meliaceae	Sentul	Stems/methanol	5 mg/ea.	BALB/c mice	significant inhibition (94%) in TPA-induced edema was observed in the isolated compound 3-oxo-12-oleanen-29-oic acid	Rasadah <i>et al.</i> , 2004
6	<i>Garcinia subelliptica</i>	Guttiferae	Pokok penanti	Seeds/chloroform	3, 10, 30, 50, and 100 μ M	Sprague-Dawley rats	A potent inhibitory effect on fMLP/CB-induced superoxide	Weng <i>et al.</i> , 2003

Table. 2 continue

S no	Scientific name	Family	Local name	Part/so lven t used	Dose of the extract	Experi mental animals	Results	Ref
7	<i>Ardisia crispa</i>	Myrsina ceae	Mata pelandok	Root/et hanol	100, and 300 mg/kg of body weigh	Sprague - Dawley rats	A significant inhibition(93.34%) was observed in carrageenan induced edema in rats at a dose of 300 mg/k	Roslida <i>et al.</i> ,20 08
8	<i>Justicia gendarussa</i>	Acantha ceae	Daun rusa	Root/ metha nol	50 mg/kg of the extract	Wistar rats	80% and 93% edema inhibition at the third and fifth hours	Kumar <i>et al.</i> ,2012
9	<i>Solanum nigrum</i>	Solanac eae	Terung meranti	Leaves /water	10, 50, and 100% of concentr ation	BALB/c mice and Sprague - Dawley rats	Extracts produce apparently two- phase anti- inflammatory activity: the first phase between 1 and 2 h	Zakaria <i>et al.</i> , 2009
10	<i>Atylosia scarabaeoides</i>	Fabaceae	kacang kerara	Leaves /etha nol	150, 300, and 450 mg/kg	Swiss albino mice	The extract displayed significant inhibition of inflammation. Highest inhibition of paw edema(38.38%) at a dose of 450 mg/kg after 4 h of administration Cucurbitacin E inhibits	Sufian <i>et al.</i> , 2017
11	<i>Kaempferia galanga</i>	Zingibe raceae	Ceku	Rhizo mes/c hlorofo rm	2 g/kg of the extract	Male Sprague - Dawley rats	Highest edema inhibition(42.9%)	Umar. <i>et al.</i> , 2012

Table 2 continue

S. no	Scientific name	Family	Local name	Part/solve n t used	Dose of theextra ct	Experimen tal animals	Results	Ref
12	<i>Stachytarph eta jamaicensis</i>	Verbenace a e	Selasih dandi	Leaves/eth ane	50, 100, and 150 mg/k g	BALB/c albino strain mice	A significant dose-dependent anti-inflammatory activity was observed 30 min after the administrati on of the extract at all doses	Sulaiman <i>et al.</i> , 2009
13	<i>Citrullus lanatus</i>	Cucurbita ceae	Tembikai	Fruit pulp/petrol eum ether, chloroform ,and 90% ethanol	30 and60 mg/k g of body weight	BALB/c mice	Cucurbitacin E inhibits inflammati on significantly from the fourth hour and is able to revert paw edema through the COX-2 inhibition	Abdel Wahabet <i>al.</i> , 2011
14	<i>Manilkara zapota</i>	Sapotaceae	Ciku	Leaves/eth ylacetate	300 mg/k g of body weight	Albino Wistar rats	Inhibitionof paw edema (92.41%)	Gangul y <i>et al.</i> , 2013
15	<i>Vitex negundo</i>	Lamiaceae	Legundi	Leaves/eth ane	2 mg/ea r	Mice	+e extract showed an inhibitory activity of 54.1%	Yunoset <i>al.</i> ,2005

Table. 2 Continue

S.no	Scientific name	Family	Local name	Part/solvent used	Dose of the extract	Experimental animals	Results	Ref
16	<i>Corchorus capsularis</i>	Malvaceae	Kancing baju	Leaves/ chloroform	20, 100, and 200 mg/kg	BALB/c mice and Sprague-Dawley rats	The extract caused significant reduction in the thickness of edematous paw for the first 6 h	Zakaria <i>et al.</i> , 2007
17	<i>Mitragyna speciosa</i>	Rubiaceae	Biak-biak and ketom	Leaves/methanol	50, 100, and 200 mg/kg	Sprague-Dawley rats	Both doses of 100 and 200 mg/kg showed a significant inhibition of the paw edema (63%)	Mosaddeq <i>et al.</i> , 2009
18	<i>Zingiber zerumbet</i>	Zingiberaceae	Lempuyang	Rhizomes/methanol	5, 10, 50, and 100 mg/kg	ICR mice	The isolated compound (zerumbone) significantly showed dose-dependent inhibition of paw edema induced by carrageenan at all doses (5, 10, 50, and 100 mg/kg) in mice with percentage of inhibition of 33.3, 66.7, 83.3, and 83.3%, respectively	Sulaiman <i>et al.</i> , 2010
19	<i>Crinum asiaticum</i>	Amaryllidaceae	Pokok bakung	Leaves/methanol	50 mg/kg of the extract	Mice	Inhibition of paw edema (94.8%)	Samud <i>et al.</i> , 1999
20	<i>Moringa oleifera</i>	Moringaceae	Kelur	Leaves/water	10, 30, and 100 mg/kg	BALB/c mice and Sprague-Dawley rat	Highest edema inhibition (66.7%) at the second hour at 100 mg/kg of dose	Sulaiman <i>et al.</i> , 2008

Table. 2 Continue

S.no	Scientific name	Family	Local name	Part/ solvent used	Dose of the extract	Experimental animals	Results	Ref
21	<i>Curcuma aeruginosa</i>	Zingiberaceae	Temuhitam	Rhizomes/ chloroform, methanol, and water	100, 200, 400, and 800 mg/kg	Swiss mice and Wistar rats	No significant suppression was observed after oral administration of all doses on carrageenan-induced paw edema	Reanmongko <i>et al.</i> , 2006
22	<i>Muntingia calabura</i>	Muntingiaceae	Kerukup siam	Leaves/water	27 mg/kg, 135 mg/kg	Sprague-Dawley rats	+e extract was found to exhibit a concentration-independent anti-inflammatory activity	Zakaria <i>et al.</i> , 2007
23	<i>Dicranopteris linearis</i>	Gleicheniaceae	Resam	Leaves/chloroform	10, 100, and 200 mg/kg	BALB/c mice and Sprague-Dawley rats	+e extract produced significant anti-inflammatory activity that did not depend on the doses of the extract	Zakaria <i>et al.</i> , 2006
24	<i>Orthosiphon stamineus</i>	Lamiaceae	Misai kucing	Leaves/methanol: water	125, 250, 500, and 1000 mg/kg	Charles River mice and Sprague-Dawley rats	Increase in edema inhibition (26.79%)	Yam <i>et al.</i> , 2008
25	<i>Curcuma longa</i>	Zingiberaceae	Kunyit	Rhizomes/water	200 mg/kg of body weight	Wistar albino rats	+e production of anti-inflammatory/proinflammatory cytokines is decreasing	Ramadhan <i>et al.</i> , 2011

Table. 2 continue

S. no	Scientific name	Family	Local name	Part/solvent used	Dose of the extract	Experimental animals	Results	Ref
26	<i>Physalis minima</i>	Solanaceae	Pokok letup-letup	Whole plant/methanol chloroform fraction	200 and 400 mg/kg	NMRI mice and Wistar rats	inhibition of paw edema at 66% and 68% at 400 mg/kg, respectively	Khan <i>et al.</i> , 2009
27	<i>Dicranopteris linearis</i>	Gleicheniaceae	Resam	Leaves/chloroform	10, 100, and 200 mg/kg	BALB/c mice and Sprague-Dawley rats	+e extract produced significant anti-inflammatory activity that did not depend on the doses of the extract	Zakaria <i>et al.</i> , 2006
28	<i>Curcuma aeruginosa</i>	Zingiberaceae	Temu hitam	Rhizomes/chloroform, methanol, and water	100, 200, 400, and 800 mg/kg	Swiss mice and Wistar rats	No significant suppression was observed after oral administration of all doses on carrageenan-induced paw edema	Reanmongko <i>et al.</i> , 2006
29	<i>Phyllanthus acidus</i>	Phyllanthaceae	Cermai	Leaves/methanol, ethyl acetate, and petroleum ether	250 and 500 mg/kg	Wistar rats and albino mice	All the extracts showed reduction in carrageenan-induced paw edema with highest inhibition (90.91%) in the methanol extract	Chakraborty <i>et al.</i> , 2012

Discussion:

According to the World Health Organization (WHO), medicinal plants are defined as those containing chemicals suitable for therapeutic use or pharmaceutical production. In developing countries, people continue to rely on these plants for treating various diseases, and the market for medicinal plant products is expanding, indicating their economic importance. Despite a vast number of plant species, only 15% of the world's 300,000 plant species have been studied for their pharmacological activity. Notably, 25% of modern medicines have origins in natural resources like medicinal plants. Recently, global attention towards researching health benefits of medicinal plants has increased, including in Malaysia, known for its diverse medicinal plant species. For example, Peninsular Malaysia and Sabah boast 1300 and 7411 documented medical plant species, respectively (Abu Bakar et al., 2018). Inflammation, a response to cell damage from pathogens or irritants, involves the activation of leukocytes and release of proinflammatory cytokines. This process includes the production of nitric oxide (NO), prostaglandin E2 (PGE2), and other mediators. Proinflammatory mediators, like nitric oxide and prostaglandins, contribute to various inflammatory conditions, and uncontrolled inflammation is associated with several chronic diseases. Acute inflammation is characterized by increased blood flow and swelling, while chronic inflammation may lead to genetic instability, neoplasia, cancer, and infectious illnesses. Nonsteroidal anti-inflammatory drugs (NSAIDs) effectively reduce COX-1 and COX-2 activity, but their use is linked to adverse effects such as gastrointestinal erosions, renal failure, and hepatic failure. COX-2 inhibitors, like Vioxx, have been associated with significant cardiovascular events. To address these issues, research has focused on anti-inflammatory medicines derived from natural sources, particularly medicinal plants abundant in phytochemicals. Enzyme inhibitory assays, such as COX and LOX, are commonly used to assess their anti-inflammatory efficacy. Tables 1 and 2 present in vitro and in vivo investigations of Malaysian medicinal plants with anti-inflammatory properties.

Conclusion:

The primary aim of this research was to examine academic studies focusing on the anti-inflammatory attributes of specific medicinal plants utilized across diverse countries. In this review, a wide array of plants from different geographical regions, including north, south, east, central, and west African countries, were identified. While various medications for treating and

managing inflammatory conditions exist in both developed and developing nations, many of these drugs are associated with undesirable side effects. Consequently, numerous investigations have been undertaken in different countries to evaluate the anti-inflammatory properties of medicinal plants, aiming to validate their utilization in traditional medicine. The exploration of plants with anti-inflammatory properties represents an emerging field in contemporary biomedicine. Given that traditional healers worldwide may possess valuable knowledge about unstudied plants, further research on those with anti-inflammatory potential is imperative.

References

1. Abas, F., Lajis, N. H., Israf, D. A., Khozirah, S., & Kalsom, Y. U. (2006). Antioxidant and nitric oxide inhibition activities of selected Malay traditional vegetables. *Food Chemistry*, 95(4), 566-573.
2. Abdelwahab, S. I., Hassan, L. E. A., Sirat, H. M., Yagi, S. M. A., Koko, W. S., Mohan, S., ... & Hadi, A. H. A. (2011). Anti-inflammatory activities of cucurbitacin E isolated from *Citrullus lanatus* var. *citroides*: role of reactive nitrogen species and cyclooxygenase enzyme inhibition. *Fitoterapia*, 82(8), 1190-1197.
3. Abdullah, Z., Hussain, K., Ismail, Z., & Ali, R. M. (2009). Anti-inflammatory activity of standardised extracts of leaves of three varieties of *Ficus deltoidea*. *International Journal of Pharmaceutical and Clinical Research*, 1(3), 100-105.
4. Abu Bakar, F. I., Abu Bakar, M. F., Abdullah, N., Endrini, S., & Rahmat, A. (2018). A review of Afsar, V., Reddy, Y. M., & Saritha, K. V. (2012). In vitro antioxidant activity and anti-inflammatory activity of methanolic leaf extract of *Boswellia serrata*. *Int J Life SC Bt & Pharm Res*, 1(4), 15-23.
5. Ana, B., Abdul, M. A., Razali, M., & Noorjahan, B. M. (2007). Anti-oxidant and anti-inflammatory activities of leaves of *Barringtonia racemosa*. *Journal of Medicinal Plants Research*, 1(5), 095-102.
6. Bandyopadhyay S, Abiodun OA, Ogboo BC, Kola-Mustapha AT, Attah EI, Edemhanria L, Chakraborty, R., Biplab, D., Devanna, N., & Sen, S. (2012). Anti-inflammatory, antinociceptive and antioxidant activities of *Phyllanthus acidus* L. extracts. *Asian pacific journal of tropical biomedicine*, 2(2), S953-S961.

7. Chan, K. Y., Mohamad, K., Ooi, A. J., Imiyabir, Z., & Chung, L. Y. (2012). Bioactivity-guided fractionation of the lipoxygenase and cyclooxygenase inhibiting constituents from *Chisocheton polyandrous* Merr. *Fitoterapia*, 83(5), 961-967.
8. Chung, L. Y., Soo, W. K., Chan, K. Y., Mustafa, M. R., Goh, S. H., & Imiyabir, Z. (2009). Lipoxygenase inhibiting activity of some Malaysian plants. *Pharmaceutical Biology*, 47(12), 1142-1148.
9. Ehsan, O., Norhani, A., Wan, Z. S., Abdul, R. O., Syahida, A., Wen, B. K., ... & Yin, W. H. (2011). Antioxidant, anti-inflammatory and anticancer activities of methanolic extracts from *Jatropha curcas* Linn. *Journal of Medicinal Plants Research*, 5(1), 49-57.
10. Foong, C. P., & Hamid, R. A. (2012). Evaluation of anti-inflammatory activities of ethanolic extract of *Annona muricata* leaves. *Revista Brasileira de Farmacognosia*, 22, 1301-1307.
11. Ganguly, A., Al Mahmud, Z., Uddin, M. M. N., & Rahman, S. A. (2013). In-vivo anti-inflammatory and anti-pyretic activities of *Manilkara zapota* leaves in albino Wistar rats. *Asian Pacific Journal of Tropical Disease*, 3(4), 301-307.
12. George, A., Chinnappan, S., Chintamaneni, M., Kotak C, V., Choudhary, Y., Kueper, T., & Radhakrishnan, A. K. (2014). Anti-inflammatory effects of *Polygonum minus* (Huds) extract (*Lineminus*) in in-vitro enzyme assays and carrageenan induced paw edema. *BMC complementary and alternative medicine*, 14(1), 1-7.
13. Hussain N, Kakoti BB, Rudrapal M, Sarwa KK, Celik I, Attah EI, Khairnar SJ, Bhattacharya S, hydro-alcoholic extract (HAE) with antioxidant potential in diabetic rats. *Indian Journal of Natural Products and Resources* 11(1): 18–29. <http://nopr.niscair.res.in/handle/123456789/54489>

14. Ibrahim, B., Sowemimo, A., van Rooyen, A., & Van de Venter, M. (2012). Anti-inflammatory, analgesic and antioxidant activities of *Cyathula prostrata* (Linn.) Blume (Amaranthaceae). *Journal of Ethnopharmacology*, 141(1), 282-289.
15. Isa, N. M., Abdelwahab, S. I., Mohan, S., Abdul, A. B., Sukari, M. A., Taha, M. M. E., ... & Mustafa, M. R. (2012). In vitro anti-inflammatory, cytotoxic and antioxidant activities of Boesenbergin A, a chalcone isolated from *Boesenbergia rotunda* (L.) (fingerroot). *Brazilian Journal of Medical and Biological Research*, 45, 524-530.
16. Journal of scientific & Technical Research 39(2): 31129–31131. <https://doi.org/10.26717/BJSTR.2021.39.006268>
17. Junejo JA, Rudrapal M, Zaman K (2020b) Antidiabetic activity of *Carallia brachiata* Lour. leaves Kanterman, J., Sade-Feldman, M., & Baniyash, M. (2012, August). New insights into chronic inflammation-induced immunosuppression. In *Seminars in cancer biology* (Vol. 22, No. 4, pp. 307-318). Academic Press.
18. Karimi, E., Jaafar, H. Z., & Ahmad, S. (2013). Antifungal, anti-inflammatory and cytotoxicity activities of three varieties of *Labisia pumila* benth: from microwave obtained extracts. *BMC Complementary and Alternative Medicine*, 13(1), 1-10.
19. Khan, M. A., Khan, H., Khan, S., Mahmood, T., Khan, P. M., & Jabar, A. (2009). Anti-inflammatory, analgesic and antipyretic activities of *Physalis minima* Linn. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 24(3), 632-637.
20. Kim, Y. H., Kim, K. H., Han, C. S., Park, S. H., Yang, H. C., Lee, B. Y., ... & Lee, N. H. (2008). Anti-inflammatory activity of *Crinum asiaticum* Linne var. *japonicum* extract and its application as a cosmeceutical ingredient. *Journal of cosmetic science*, 59(5), 419-430.
21. Kumar R (2021) Epidemiology of Life-Threatening Disease and Inflammation. Biomedical Kumar, K. S., Vijayan, V., Bhaskar, S., Krishnan, K., Shalini, V., & Helen, A. (2012). Anti-inflammatory potential of an ethyl acetate fraction isolated from *Justicia gendarussa* roots through inhibition of

iNOS and COX-2 expression via NF- κ B pathway. *Cellular Immunology*, 272(2), 283-289.

22. Kumari A, Jaganathan R, Adalakun NS (2021) Polypharmacology of some medicinal plant metabolites against SARS-CoV-2 and host targets: Molecular dynamics evaluation of NSP9 RNA binding protein. *Journal of Biomolecular Structure and Dynamics* 25: 1–7 <https://doi.org/10.1080/07391102.2021.1959401>

23. Kundu, J. K., & Surh, Y. J. (2012). Emerging avenues linking inflammation and cancer. *Free Radical Biology and Medicine*, 52(9), 2013-2037.

24. Kutoi, C. J., Yen, K. H., & Seruji, N. M. U. (2012, April). Pharmacology evaluation of *Litsea garciae* (Lauraceae). In 2012 IEEE Business, Engineering & Industrial Applications Colloquium (BEIAC) (pp. 31-33). IEEE.

25. Lahare RP, Bisen YK, Yadav HS, Dashahre AK (2021) Estimation of Total Phenol, Flavonoid, Lee, K. H., Padzil, A. M., Syahida, A., Abdullah, N., Zuhainis, S. W., Maziah, M., ... & Lajis, N.H. (2011). Evaluation of anti-inflammatory, antioxidant and antinociceptive activities of six Malaysian medicinal plants. *J Med Plant Res*, 5(23), 5555-63.

26. Liu, J. Z., Zhang, C. C., Fu, Y. J., & Cui, Q. (2022). Comparative analysis of phytochemical Malaysian medicinal plants with potential anti-inflammatory activity. *Advances in pharmacological sciences*, 2018.

27. Mogana, R., Teng-Jin, K., & Wiart, C. (2013). Anti-inflammatory, anticholinesterase, and antioxidant potential of Scopoletin isolated from *Canarium patentinervium* Miq. (Burseraceae Kunth). *Evidence-based complementary and alternative medicine*, 2013.

28. Mosaddeq, W. S., Sulaiman, M. R., Mohamad, T. T., Chiong, H. S., Zakaria, Z. A., Jabit, M. L., ... & Israf, D. A. (2009). Anti-inflammatory and antinociceptive effects of *Mitragyna speciosa* Korth methanolic extract. *Medical Principles and Practice*, 18(5), 378-384. profile, antioxidant and anti-inflammatory activity from *Hibiscus manihot* L. flower. *Arabian Journal of Chemistry*, 15(1), 103503.

29. Ramadan, G., Al-Kahtani, M. A., & El-Sayed, W. M. (2011). Anti-

inflammatory and anti-oxidant properties of *Curcuma longa* (turmeric) versus *Zingiber officinale* (ginger) rhizomes in rat adjuvant-induced arthritis. *Inflammation*, 34(4), 291-301.

30. Ramsewak, R. S., DeWitt, D. L., & Nair, M. G. (2000). Cytotoxicity, antioxidant and anti-inflammatory activities of curcumins I–III from *Curcuma longa*. *Phytomedicine*, 7(4), 303-308.

31. Rasadah, M. A., Khozirah, S., Aznie, A. A., & Nik, M. M. (2004). Anti-inflammatory agents from *Sandoricum koetjape* Merr. *Phytomedicine*, 11(2-3), 261-263.

32. Reanmongkol, W., Subhadhirasakul, S., Khaisombat, N., Fuengnawakit, P., Jantasila, S., & Khamjun, A. (2006). Investigation the antinociceptive, antipyretic and anti-inflammatory activities of *Curcuma aeruginosa* Roxb. extracts in experimental animals. *Songklanakarini J Sci Technol*, 28(5), 999-1008.

33. Roslida, A., & Kim, K. (2008). Anti-inflammatory and anti-hyperalgesia effects of *Ardisia crispa* Thunb. DC. *Pharmacognosy Magazine*, 4(16), 262.

34. Sahoo RK, Walode SG (2021) Bioactive Antidiabetic Flavonoids from the Stem Bark of *Cordia dichotoma* Forst.: Identification, Docking and ADMET Studies. *Molbank* 2021(2):M1234. <https://doi.org/10.3390/M1234>

35. Samud, A. M., Asmawi, M. Z., Sharma, J. N., & Yusof, A. P. M. (1999). Anti-inflammatory activity of *Crinum asiaticum* plant and its effect on bradykinin-induced contractions on isolated uterus. *Immunopharmacology*, 43(2-3), 311-316.

36. Shaari, K., Suppaiah, V., Wai, L. K., Stanslas, J., Tejo, B. A., Israf, D. A., ... & Lajis, N. H. (2011). Bioassay-guided identification of an anti-inflammatory prenylated acylphloroglucinol from *Melicope ptelefolia* and molecular insights into its interaction with 5-lipoxygenase. *Bioorganic & medicinal chemistry*, 19(21), 6340-6347. signaling pathways and targeted intervention. *Signal Transduction and Targeted Therapy* 6(1): 1–46. <https://doi.org/10.1038/s41392-021-00658-5>

37. Siriwatanametanon, N., Fiebich, B. L., Efferth, T., Prieto, J. M., & Heinrich, M. (2010).

Traditionally used Thai medicinal plants: in vitro anti-inflammatory, anticancer and antioxidant activities. *Journal of ethnopharmacology*, 130(2), 196-207.

38. Sufian, M. A., Islam, M. R., Chowdhury, T. K., Rahman, A., Uddin, M. S., Koly, S. F., & Sarwar, M. S. (2017). Investigation of in vivo analgesic, anti-inflammatory, in vitro membrane stabilizing and thrombolytic activities of *Atylosia scarabaeoides* and *Crotalaria specialist* leaves. *Journal of Pharmacology and Toxicology*, 12(3), 120-128.
39. Sulaiman, M. R., Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Tasrip, N. A., ...& Israf, D. A. (2010). Anti-inflammatory effect of zerumbone on acute and chronic inflammation models in mice. *Fitoterapia*, 81(7), 855-858.
40. Sulaiman, M. R., Zakaria, Z. A., Bujarimin, A. S., Somchit, M. N., Israf, D. A., & Moin, S. (2008). Evaluation of *Moringa oleifera* aqueous extract for antinociceptive and anti-inflammatory activities in animal models. *Pharmaceutical biology*, 46(12), 838-845.
41. Sulaiman, M. R., Zakaria, Z. A., Chiong, H. S., Lai, S. K., Israf, D. A., & Shah, T. A. (2009). Antinociceptive and anti-inflammatory effects of *Stachytarpheta jamaicensis* (L.) Vahl (Verbenaceae) in experimental animal models. *Medical Principles and Practice*, 18(4), 272-279.
42. Tannin and Alkaloid Content in Different Extracts of *Catharanthus roseus* from Durg District, Chhattisgarh, India. *Scholars Bulletin* 7(1): 1–6. <https://doi.org/10.36348/sb.2021.v07i01.001>
43. Umar, M. I., Asmawi, M. Z., Sadikun, A., Atangwho, I. J., Yam, M. F., Altaf, R., & Ahmed, A. (2012). Bioactivity-guided isolation of ethyl-p-methoxycinnamate, an anti-inflammatory constituent, from *Kaempferia galanga* L. extracts. *Molecules*, 17(7), 8720-8734.
44. Varghese, C. P., Ambrose, C., Jin, S. C., Lim, Y. J., & Keisaban, T. (2013). Antioxidant and anti-inflammatory activity of *Eurycoma longifolia* Jack, a traditional medicinal plant in Malaysia. *Int J Pharm Sci Nanotechnol*, 5(4), 1875-8.
45. Vijaya Kumar, S., Sankar, P., & Varatharajan, R. (2009). Anti-inflammatory activity of roots of *Achyranthes aspera*. *Pharmaceutical Biology*, 47(10), 973-975.
46. Weng, J. R., Lin, C. N., Tsao, L. T., & Wang, J. P. (2003). Novel and anti-inflammatory constituents of *Garcinia subelliptica*. *Chemistry—A European Journal*, 9(9), 1958-1963.

47. Yam, M. F., Asmawi, M. Z., & Basir, R. (2008). An investigation of the anti-inflammatory and analgesic effects of *Orthosiphon stamineus* leaf extract. *Journal of medicinal food*, 11(2),362-368.
48. Yunos, N. M., Kean, O. B., & Abas, R. (2005). Cytotoxicity evaluations on *Vitex negundo* anti-inflammatory extracts. *Malaysian Journal of Science*, 24(1), 213-217.
49. Zakaria, Z. A., Ghani, Z. D. F. A., Nor, R. N. S. R. M., Gopalan, H. K., Sulaiman, M. R., & Abdullah, F. C. (2006). Antinociceptive and anti-inflammatory activities of *Dicranopterislinearis* leaves chloroform extract in experimental animals. *Yakugaku Zasshi*, 126(11),1197-1203.
50. Zakaria, Z. A., Hussain, M. K., Mohamad, A. S., Abdullah, F. C., & Sulaiman, M. R. (2012). Anti-inflammatory activity of the aqueous extract of *Ficus deltoidea*. *Biological research for nursing*, 14(1), 90-97.
51. Zakaria, Z. A., Nor Hazalin, N. A., Zaid, S. N. H., Ghani, M. A., Hassan, M. H., Gopalan, H. K., & Sulaiman, M. R. (2007). Antinociceptive, anti-inflammatory and antipyretic effects of *Muntingia calabura* aqueous extract in animal models. *Journal of Natural Medicines*, 61(4),443-448.
52. Zakaria, Z. A., Sulaiman, M. R., Gopalan, H. K., Ghani, Z. D. F. A., Nor, R. N. S. R. M., Jais, A.M. M., & Abdullah, F. C. (2007). Antinociceptive and anti-inflammatory properties of *Corchorus capsularis* leaves chloroform extract in experimental animal models. *YakugakuZasshi*, 127(2), 359-365.
53. Zakaria, Z. A., Sulaiman, M. R., Morsid, N. A., Aris, A., Zainal, H., Mohd Pojan, N. H., & HananKumar, G. (2009). Antinociceptive, anti-inflammatory and antipyretic effects of *Solanumnigrum* aqueous extract in animal models. *Methods and findings in experimental and clinical pharmacology*, 31(2), 81.