Effects of *Moringaoleifera* leaf extract on the carcass traits, growth and physiological biomarkers of broiler chicken.

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Abstract: Aspire of the present study was to scrutinize the effects of *Moringaoleifera*leaf extract (MOLE) on various growth biomarkers (feed intake,final live weight,weight gain and feed conversion ratio)of broiler chicks. A 6 weeks trials study was done on Day old broiler chicks (Hubbard) with 350 in number distributed into 5 equal groups. Group 1st were named as Control group was fed on basal feed common to other groups and water without MOLE. The othertreatment groups MOLE₂, MOLE₄, MOLE₆ andMOLE₈ were provided with the basal feed with2, 4, 6 and 8mL of*Moringaoleifera*leaf extract per liter of their drinking water respectively. It was observed through the results that the broilers chicks provided with4% MOLEin watershowed significant (*P*<0.05) highestfinal and net body weight in comparison to control and other treatment groups. A significant (*P*<0.05) increased trend in live and

I.INTRODUCTION

Poultry production is a mean of earnings; that is an excellent protein source and immediate income on investment [40, 41]. Poultry production represents a significant pillar of food safety measures development. Similarly it plays a key role in socio-economic and cultural development in various countries [1, 3, 12]. But there are few confronts faced by the emergent countries; which comprise ratio of high feed to gain and expensive feed ingredients that are becoming costly day by day. A number of efforts have been done to trounce these confronts which one includes the providing of antibiotics through nutrition [1, 9, 20, 44].

The antibiotics are being used as growth promoters and as preventive measures for diseases [18, 25, 31]. Beyond using antibiotics as growth enhancer, these have few drawbacks including residual effects, drug toxicity and rise in bacterial resistance [24, 23, 18, 33]. Such ills have guided the European Union to prohibit antibiotics use like growth promoters [8]. Therefore, awareness to natural plus safe choices such as herbs alternate to antibiotics is increasing day by day [15, 42].

Moringaoleifera is a well renowned easily cultivable plant. The treeof *M. oleifera* is indigenous to South Asia particularly Pakistan, Bangladesh, India, Afghanistan, Sri Lanka, Africa,

dressed body weight was observed as compared to control group. As compared to control group, significant increased values of Hemoglobin and Packed cell volume were observed in broiler chicks fed on 2, 4,6, 8% of MOLE. Same trend was observed for white blood cells. However it was observed through the experiment that chicken of control group had lower counts of RBC and WBC, PCV and lymphocyte levels. It was concluded in the study that adding of 4% MOLE in drinking water caused an increase in growth performance and carcass traits. The broiler fed on feed with 4% Moringa showed highest performance compared to others .

Keywords: Broiler, *Moringaoleifera* leaf extract, Physiological biomarkers, Carcass traits, growth

Arabia and Madagascar [27, 30, 43]. Being enriched with nutrition due to existence of a diversity of essential phytochemicals, the various segments of the plant are utilized like a fine resource of nutrition for human being and in conventional food in various regions of the globe [39, 44]. There is a broad use of different segments of moringa in the therapeutics and feed industry [12, 13]. It is well-liked for its leaves, seeds, pods and flowers in human feed and herbal medicine [6, 7, 36].

Though, the data regarding the effects of extract of leaf meal of the plant in the feed on growth and meat quality of broiler is incomplete and differing. [28] and [4] concluded a reduced ingestion however enhance in feed conversion ratio (FCR). It was reported that when the concentration of *Moringa oleifera* leaf meal content was beyond 5% in the feed, a decrease in growth graph of broiler chickens was observed [19, 26]. [21] and [37] approved it the using *Moringa oleifera* leaf meal to 10% level had no adverse effect on the prolific act of broiler chicken, on the other hand higher than 10 % cause unhealthy things. It might be due to impure extraction of leaf meal, cooked or tinctured as a leaf extract [17]. Additionally, the poor production at increased constituent level might be owing to increased levels of antinutritional factors, poor digestibility of the energy fiber [16], dustiness and occurrence of protein in the raw leaf [1, 14].

II. METHODOLOGY

Study location:

The current study was conducted at postgraduate laboratory of Faculty of Veterinary & Animal Sciences, Gomal University Dera Ismail Khan.

Test ingredients and preparation of the extracts:

Fresh *Moringa oleifera* leaves were obtained from the local farmers. The leaves were washed thoroughly and dried in air by spread on a polythene paper sheeth in shady zone. The dried leaves were ground into powder using an electric grinder at department of Animal and Poultry production, Gomal University Dera Ismail Khan. 100 g of the powder was then soaked for 24 hrs in 01 liter of absolute fresh water and was shaken regularly. Using a muslin cloth, the preparation was then filtered to remove the remains. The extractwas kept in dirt free air tight container at 4°C for further use.Aqueous extractof different concentrations 2mL, 4mL, 6mL and 8mL was made by *M.oleifera*.

The body weight was measured at the start of experiment. Then at the end of each week, the body weight was recorded. Feed intake was measured weekly. Feed conversion ratio (FCR) was determined based on feed consumption and body weight gain.

Housing management:

After washing with detergent and water, the walls and floors of the bird keeping units were disinfected. The wood shavings were used as litter materials and old newspapers spread on for 1st one week of the chicks. Dry, disinfected and clean drinkers and feeders were set in a position reachable to the chicks. Proper labeling of each pen was done for easy recognition of groups. At start of 02 weeks, Super starter mash was given to the birds and starter mash throughout the third and fourth week. At the fifth week of age, finisher feed pellets was given till the sixth week. The respective feeds were offered *ad libitum* and changing one type of feed to next was done slowly to avoid digestive problems. Proper medications as well as vaccinations were done also to the birds.

Experimental Design:

The 350 birds were divided into five groups as follows;C;Control group fed on conventional feed plus fresh drinking water, "MOLE₂" groupfed on conventional feed with *M. oleifera* leaf extract at the rate of 2mL/L, "MOLE₄" added with *M. oleifera* leaf extract at the rate of 4mL/L. "MOLE₆" with *M. oleifera* leaf extract at the rate of 6mL/L, "MOLE₈" with *M. oleifera* leaf extract at the rate of 8mL/L.

Carcass traits and blood analysis:

At the end of experiment; 5 birds selected randomly from each group was taken and were fasted overnight and slaughtered. The dressing percentage was calculated by live body weight and dressed after removal of viscera, feet and skull.

Total 3ml blood was collected in EDTA coated vaccutainers for further analysis. Complete blood count (CBC) was determined using hematology analyzer.

Statistical Analysis:

Data was analysed through one way ANOVA using statistical package SPSS 21. Association between variables was considered statistically significant at P<0.05. Tukey's test was applied to compare the means.

III. RESULTS

Different birds of various treatment groups of C, MOLE₂, MOLE₄, MOLE₆ and MOLE₈ consumed the feed 4986.54 ± 10.3 , 5062.13±12.4, 5048.9±10.45 4852.23±11.2. and 4887.21±11.6g/bird respectively. No significant difference was observed among the groups. Table No. 1 showing the summary of final body weight, total weight gain and FCR of different treatment groups. It was observed that the birds supplemented with 4% Moringa aqueous extract have the highest body weight as compared to other groups. On the other hand the birds treated with 8% MOLE were with the lowerfinal body weight.Similar pattern wasobserved for daily weight gain that was higher in the group MOLE₄ as compared to birds of other groups.FCR was significantly better in group MOLE₄. The group MOLE₄ showed the lowest FCR ratio as compared to other treatment groups including control group.

A significant increase in dressing weight and % age was observed. The birds of group MOLE₄ showed significant increased dressed weight (Table No.2). No significant difference (P<0.05) was observed in different treatment for dressing percentage. A significant decline in relative weight of proventriculus was observed but non-significant difference was noted in relative weight of heart, gizzard and spleen of birds fed on MOLE.

Physiological parameters:

Blood profile is biomarker of an animal for nutritional, physiological and pathological status. The effect of MOLE on different blood parameters of treatment groups is shown in (Table No.3). As compared to control group, significant (P<0.05) increased values of Hemoglobin and Packed cell volume were observed in broiler chicks fed on 2, 4,6, 8% MOLE. Same trend was observed for white blood cells. Significant (P<0.05) increased values of WBC were observed for all the treatment groups as compared to control group while a non significant difference was observed for RBCs.

IV. DISCUSSION

Feed intake, Body weight gain and FCR ratio were evaluated and implicated as key markers for growth performance of experimental chickens. The results of current study highlighted that significant increased concentration of *Moringaoleifera* significantly enhanced the water intake in the treatment groups MOLE2, MOLE4 but decreased in MOLE6 and MOLE8. The findings in the study are in parallel to the conclusions made by [11, 5, 29]. The results showed that significant increased concentrations significantly enhanced the feed intake. As leaves of the plant are enriched naturally with vitamins, minerals and proteins especially 8 essential amino acids [24]. The increased feed intake with increased concentration of *Moringa oleifera* leaves extract might be due to its maximum digestibility and lack of any factor that reduce the feed intake [45].

It was observed through the current study that 4% of leaf extract supplemented in feed enhanced the gain in body weight. Concentrations higher than 4% declined the graph. Same observations were observed by a most recent research [32]. [17] also endorsed the conclusion but opposed by [10] and [47]. Hence it is still questionable that why MOLE impacts negatively on weight gain on higher concentrations. It might be due to presence of some anti-nutritional factors to an extent to impair the digestibility ultimately inhibiting the growth of broiler. These substances may be the tannins [48] which impede the nutritional digestion which consequences in slowing down of absorption and metabolic activities [32]. In another study conducted previously by [22] that tannins may interfere the growth of broiler negatively when their concentration becomes higher than 5g/kg of feed. FCR was significantly better in treatment group "MOLE4" supplemented with MOLE at the rate of 4mL/L of drinking water.

The treatment group MOLE4 showed the lowest FCR ratio as

compared to other treatment groups including control group "C". The calculations found in current study are endorsed by a most recent study [32] opposite to the findings of [2].

Blood plays a crucial role in physiology of broiler poultry [31]. In our current study it was revealed that comparing the control group, significant increased values of Hemoglobin and Packed cell volume were observed in broiler chicks provided with 2, 4,6, 8% MOLE. Same trend was observed for white blood cells [35] but there was no significant difference in Packed Cell Volume (%PCV), levels of White Blood Cell (WBC) count and Red Blood cell (RBC) counts among the experimental broiler chicks [38]. As observed in current study that blood hematological values were in the range of reference values. Thus *Moringaoleifera*does not contain such anti-nutritional factors which may potentiate the threat to growth of broiler. The findings of current study are strongly endorsed by [15].

 Table No.1 Growth performance of broilers at different experimental diets

Item	Groups						
	С	MOLE ₂	MOLE ₄	MOLE ₆	MOLE ₈		
Water Intake (L)	8648±15.47 ^a	8512±14.44 ^a	8961±15.48 ^b	9018±13.38 ^b	8316±14.19°		
Initial body weight(g)	50.14±0.51ª	51.88±0.76ª	50.11±0.63ª	52.17±0.42 ^a	49.58±0.88ª		
Final body weight (g)	2276±9.31ª	2367±10.12 ^b	2492±11.26°	2403±12.16 ^{bc}	2041±14.16 ^a		
Weight gain (g)	2242±9.39ª	2305±9.01ª	2517±10.52 ^b	2382±15.06 ^{ab}	1983.12°		
Feed intake	4986.54±10.3ª	4852.2±11.2ª	5062.13±12.4 ^b	5048.9±10.45 ^b	4887.21±11.6ª		
FCR	2.15±0.001 ^b	2.01±0.002 ^a	1.85±0.001°	2.06±006ª	2.38±0.003 ^b		

*Values in rows having dissimilarity in superscripts (a, b & c) are significantly (P < 0.05) differ

Characteristic	Groups					
	С	MOLM ₂	MOLM ₄	MOLM ₆	MOLM ₈	_
Live body weight	2272±9.21	2345±9.62	2478±10.56	2248±13.56	2135±14.96	0.036
Dressed weight	1797±10.51	1927±9.16	1964±10.26	1629±12.92	1445±14.67	0.000
Dressing %	79.09±0.52	82.17±2.87	79.25±1.62	72.46±1.26	67.68±0.89	0.002

Table No.2 Carcass characteristics of different groups

Table No.3 Hematology profile of broilers of different treatments groups

Parameter	Groups						
-	С	MOLE ₂	MOLE ₄	MOLE ₆	MOLE ₈		
Hb (g/dL)	12.65±1.61ª	11.47±1.32ª	11.29±1.85ª	11.22±1.07 ^a	10.12±1.41 ^b		
RBC (×10 ¹² /L)	$2.18{\pm}1.02^{a}$	2.24 ± 1.29^{a}	2.18±1.01 ^a	2.13±1.12 ^a	$2.01{\pm}1.08^{a}$		
WBC(×10 ⁹ /L)	$5.38{\pm}1.42^{a}$	5.59±1.38 °	$5.88{\pm}1.61^{a}$	5.98±1.72 ^a	6.48±1.93 ^a		
PCV (%)	38.93±3.16 ^a	36.64±2.21 ^a	36.98±2.25 °	35.02±2.04 ^a	32.15±2.26 ^b		
MCV (fL)	143.73±12.54ª	$140.54{\pm}12.24^{b}$	141.23±12.48 ^a	143.53±13.01 ^a	144.13±14.23 ^a		
MCH (pg)	40.18±2.13ª	41.12±2.07 ^a	41.23±2.51 ^a	39.78±2.84 ª	40.89±2.86 ª		
MCHC (g/dL)	28.35±2.82 ^a	28.46±2.11 ^a	28.93±2.35 ª	29.11±2.13 ^a	29.76±2.82 ^a		

*Values presented as Mean \pm Standard Error Mean(SEM). The mean in rows having differ superscripts are dissimilar to each other significantly (*P*<0.05).

REFERENCES:

- 1. Abbas, T.E. 2013. The use of *Moringa oleifera* in poultry diets. Turkish J. Vet Anim Sci., 37: 492–496.
- Abou Sekken, M.S.M. Performance, immune response and carcass quality of broilers fed low protein diets contained either Moringa oleifera leaves meal or its extract. J. Am. Sci. 2015, 11, 153–164.
- Advisory, E., & Down, C. (2019). Poultry sector study. <u>http://www.bdsknowledge.org/</u> dyn/bds/docs/960/DFID%20Poultry%20Sector%20Study% 20180419.pdf
- 4. Agbogidi, O., Ilondu, E., 2012. *Moringa oleifera* Lam: its potentials as a food security and rural medicinal item. J. Bio. Innov. 1: 156–167.
- Alnidawi, A., Ali, F., Abdelgayed, S., Ahmed, F., Farid, M. 2016. *Moringa oleifera* leaves in broiler diets Effect on chicken performance and health. Food Sci. Quality Manag. 58, 40–48.

- Anwar, F., Rashid, U., 2007. Physicochemical characteristics of *Moringa oleifera* seeds and seed oil from a wild provenance of Pakistan. Pak. J. Botany 39: 1443–1453.
- Anwar, F., Sajid, L., Muhammad, A., Anwarul, H.G., 2007. *Moringa oleifera:* A food plant with multiple medicinal uses. Phytother. Res. 21: 17–25.
- Anwar, M.I., Muhammad, F., Awais, M.M., Akhtar, M. 2017. A review of glucans as a growth promoter and antibiotic alternative against enteric pathogens in poultry. World's Poult. Sci. J. 73: 651–661.
- Arora, D.S., Onsare, J.G., Kaur, H. 2013. Bioprospecting of Moringa (Moringaceae): microbiological perspective. J. Pharmacogn. Phytochem. 1: 193–215.
- Balarabe, S., Jibir, M., Duru, S. and Abdu1, S.B. 2021. Evaluation of moringa (*Moringa oleifera*) leaf meal for broiler chicken performance and meat quality. ADAN J. Agric. 2 (1): 189-201
- 11. Baqer H. Q., Ibrahim B.M. 2022. Effect of adding pomegranate (*Punica granatum*) and moringa (*Moringa*

oleifera) in productive performance, relative growth rate, water consumption and production index of broiler chickens. J. Kerbala Agri. Sci. 3(9) 1-16.

- Chang, H. S. (2007). Analysis of the Philippine chicken industry. Asian Journal of Agriculture and development. 4(1): 1.
- Cheng, Y., Chen, Y., Li, J., Qu, H., Zhao, Y., Wen, C., Zhou, Y. 2019. Dietary Sitosterol improves growthperformance, meat quality, antioxidant status, and mitochondrial biogenesis of breast muscle in broilers. Animals. 9,71.
- Chuang, P.H., Lee, C.W., Chou, J.Y., Murugan, M., Shieh, B.J., Chen, H.M. 2007. Antifungal activity of crude extracts and essential oil of *Moringa oleifera* Lam. Bioresour.Technol. 98: 232–236.
- 15. Djouhou, M. C. F., Nwaga, D., Mafogang, B. and Fokou, E. (2020). Inclusion of *Moringa oleifera* Leaf Powder in Broiler Chicken Feed and its Effect on Growth Performance, Biochemical Profile and Metabolism Stimulation: Case Study of the Synthesis of Some Sulphur Compounds. *Biomedical Journal of Science and Technological Research* 28 (4). DOI:10.26717/BJSTR.2020.28.004693.
- 16. Fahey, J.W. (2005). *Moringa oleifera*: a review of the medical evidence for its nutritional, therapeutic and prophylactic properties. Trees Life J. 1: 1–15.
- 17. Gadzirayi CT and Mupangwa JF (2014). Feed intake and growth performance of indigenous chicks fed diets with Moringaoleifera leaf meal as a protein supplement during early brooding stage. Int. J. Poult. Sci. 13:145-150.
- Gopalakrishnan, L., Doriya, K., Kumar, D.S. 2016. *Moringa oleifera*: A review on nutritive importance and its medicinal application. Food Sci. Human Wellness 5: 49–56.
- Gupta, A., Gautam, M.K., Singh, R.K., Kumar, M.V., Rao, C.V., Goel, R.K., Anupurba, S. 2010. Immunomodulatory effect of Moringa oleifera Lam. extract on cyclophosphamide induced toxicity in mice. Indian J. Exp. Biol. 48: 1157–1160.
- 20. Gupta, S., Jain, R., Kachhwaha, S., Kothari, S.L. 2018. Nutritional and medicinal applications of Moringa oleifera Lam-Review of current status and future possibilities .J. Herbal Med.11: 1–11.
- Jaiswal, D., Kumar, P.R., Chatterji, S.M.S., Shukla, S., Rai, D.K., Sharma, G., Sharma, B., khair, S., Watal, G. 2013. Role of Moringa oleifera in regulation of diabetes-induced oxidative stress. Asian Pacific J. Tropic. Med. 10: 426–432.
- Jamroz, D., Wiliczkiewicz, A., Wertelecki, T., Orda, J., Skorupinska, J.. (2009). Use of active substances of herbal origin in chicken diets based on maize and domestic grains. *Britian Journal of Poultry Science*, 46, 485–493.
- 23. Jung, I.L. 2014. Soluble extract from Moringa oleifera leaves with a new anticancer activity. PLoS ONE, 9:954-92.
- Leone, A., Spada, A., Battezzati, A., Schiraldi, A., Aristil, J., Bertoli, S. 2015. Cultivation, genetic, ethnopharmacology,phytochemistry and pharmacology of Moringa oleifera leaves: An overview. Int. J. Mol. Sci. 16: 12791–12835.
- 25. Luqman, S., Srivastava, S., Kumar, R. Maurya, A.K., Chanda, D. 2012.Experimental assessment of Moringa

oleifera leaf and fruit for its antistress, antioxidant and scavenging potential using in vitro and in vivo assays. Evi.Bas.Compl.Alt. Med. 519084, 12.

- Mahanta, J.D., Borgohain, B., Sarma, M., Sapcota, D., Hussain, J. 2017. Effect of dietary supplementation of herbal growth promoter on performance of commercial broiler chicken. Indian J. Anim. Resh. 51: 1097–1100.
- Maheshwari, K., Yadav, R.K., Malhotra, J., Dhawan, N.G., Mohan, L. 2014. Fascinating nutritional, prophylactic, therapeutic and socio-economic reconcile attributable to drum stick tree (Moringa oleifera Lam.). Glob. J. Med. Res. B: Pharm. Drug Discov.Toxicol. Med., 14: 11–22.
- Mahfuz, S, Song, H., Miao, Y., Liu, Z. 2018. Dietary inclusion of mushroom (Flammulinavelutipes) stem waste on growth performance and immune responses in growing layer hens. J. Sci. Food Agric. 99: 703–710.
- 29. Mahfuz, S., Song, H., Wei, J., Chen, M., Zhen, D., Nahar, Z., Liu, Z.2018. Organic egg production, egg quality, calcium utilization, and digestibility in laying hens fed with mushroom (Flammulinavelutipes) stem waste.Brazilian J. Poult. Sci. 20: 717–724.
- Mahfuz, S.U., Chen, M., Zhou, J.S., Wang, S., Wei, J., Liu, Z., Song, H. 2018. Evaluation of golden needle mushroom(Flammulinavelutipes) stem waste on pullet performance and immune response. South African J. Anim. Sci, 48: 563–571.
- Makama, R.S., Duru, S., Onimisi, P.A., Moses, O. (2020). Growth performance and hematological parameters of broiler chicken fed raw and boiled senna obtusifolia seed meal. *Nigerian Journal of Animal Science*, 22(3), 271– 278.https://www.ajol.info/index.php/tjas/article/view/20214 6.
- **32.** Marifah, B, Agusetyaningsih, I., Sarjana, T., Kismiati, S., Sugiharto, S. 2023. Effect of *Moringa oleifera* Leaves Extract, Whey Protein, and Their Combination on Growth, Carcass and Meat Quality of Broiler Chickens. Tropical Ani. Sci. J. 46(3):313-320
- 33. Movahhedkhah, S., Rasouli, B., Seidavi, A., Mazzei, D., Laudadio, V., Tufarelli, V. 2019. Summer savory (Saturejahortensis L.) extract as natural feed additive in broilers: Effects on growth, plasma constituents, immune response, and ileal microflora. Animals 9, 87.
- Moyo, B., Masika, P.J., Muchenje, V., 2016. Potential use of Moringa oleifera leaf in animal feeding: A Review. Int. J. Current Agric. Res, 4: 9187–9194.
- Moyo, S.; Jaja, I.F.; Mopipi, K.; Masika, P.; Muchenje, V. 2021.Effect of graded levels of Imbrasia belina meal on blood lipid profile, bone morphometric and mineral content of broiler chickens. Anim. Feed. Sci. Technol. 271, 114736.
- 36. Obadire, F. O., Obadire, S. O., Ige, I.P., Osofowora, A. O. Oke, C.O. 2023. Growth Performance And Blood Indices Of Broiler Chickens Fed Lye-Processed Moringa (*Moringa oleifera*) seed meal *FUDMA* Journal of Agriculture and Agricultural Technology ISSN: 2504-9496 9:1-11.
- Ogbunugafor, H.A., Eeneh, F.U., Ozumba, A.N., Igwoezikpe, M.N., Okpuzor, J., Igwilo, I.O., Adenekan, S.O., Onyekwelu, O.A. 2011. Physico-chemical and anti-oxidant

properties of Moringa oleifera seed oil. Pak. J. Nutri, 10: 409-414.

- Oluduro, A.O. 2012. Evaluation of antimicrobial properties and nutritional potentials of Moringa oleifera Lam. Leafin South-Western Nigeria.J. Microbiol. 8: 59–67.
- Oluduro, O.A., Aderiye, B.I., Connolly, J.D., Akintayo, E.T., Famurewa, O. 2010. Characterization and antimicrobialactivity of 4-(_-D-glucopyranosyl-1!4-(-Lrhamnopyranosyloxy)-benzyl thiocarboxamide; a novel bioactivecompound from Moringaoleifera seed extract. Folia Microbiol. 55: 422–426.
- 40. Olugbemi, T.S., Mutayoba, S.K., Lekule, F.P. 2010. Effect of Moringa (Moringa oleifera) inclusion in cassava based diets fed to broiler chickens. Int. J. Poult. Sci. 9: 363–367.
- 41. Onunkwo, D.N., George, O.S. 2015. Effects of Moringa oleifera leaf meal on the growth performance and carcass characteristics of broiler birds. J. Agric. Vet. Sci. 8: 63–66.
- 42. Qwele, K., Hugo, A., Oyedemi, S.O., Moyo, B., Masika, P.J., Muchenje, V. 2013. Chemical composition, fatty acid content and antioxidant potential of meat from goats supplemented with moringa (Moringa oleifera) leaves, sunflower cake and grass hay. Meat Sci. 93: 455–462.
- 43. Rachmawati, I., Rifai, M. 2014. In vitro immunomodulatory activity of aqueous extract of Moringa oleifera Lam.eaf to the CD4+, CD8+ and B220+ cells in Mus musculus. J. Exp. Life Sci., 4: 15–20.
- 44. Stohs, S.J., Hartman, M.J. 2015. Review of the safety and efficacy of Moringa oleifera. Phytother. Res. 29: 796–804.
- 45. Teteh A, Voemesse K, Agbonon, Gbeassor M, Decuypere E, Tona K (2017). Effect of MoringaOleifera Leaves on transit and morphometric parameters of the digestive tract of layer pullets and laying hens. Eur.Poult.Sci. 81.10.1399/eps.173.
- Tiloke, C., Phulukdaree, A., Chuturgoon, A.A. 2013. The antiproliferative effect of Moringa oleifera crude aqueous leaf extract on cancerous human alveolar epithelial cells. BMC. Complement. Altern. Med, 13: 226–233.
- 47. Ullah, F., M. Tahir, S. Naz, N. A. Khan, & R. U. Khan. 2022. *In vitro* efficacy and ameliorating effect of *Moringa oleifera* on growth, carcass, stress and digestibility of nutrients in *Escherichia coli*-infected broilers. J. Appl. Anim. Res. 50:118-124.
- Wahyuni, R., W. Wignyanto, S. Wijana, & S. Sucipto. (2020). Optimization of protein and tannin extraction in *Moringa oleifera leaf* as antioxidant source. *Food Research*. *4*, 2224–2232.