

## Mutation Breeding Strategies for High Energy Tree Species *Prosopis juliflora* in Pakistan

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### Abstract

Pakistan is facing a severe energy crisis that is negatively impacting the whole economy as this deficiency forced local industries to move to neighboring countries. This also affects the local people losing their jobs. Due to the energy crisis, Pakistanis are forced to use tree biomass for their energy uses. This resulted in deforestation and desertification causing climate change. To combat these issues, increased afforestation of marginal and desert lands is important to meet the local demands. *Prosopis juliflora* is the main species of the desert where it is grown for multiple uses. This study explores mutation breeding's potential to develop useful mutants of *Prosopis* species. *Prosopis* species are characterized by low biomass, poor tree shape, small diameter, and long needles hinder their widespread adoption. Two types of mutagens i.e., physical (gamma radiation) and chemical (Ethyl methanesulfonate) were used for mutagenesis in *P. juliflora* and *P. cineraria* species. Seeds of both species were treated with three levels of mutagens i.e., low (15mM of EMS, 600Gy of gamma rays), moderate (25mM of EMS, 700Gy of gamma rays), and high (35mM of EMS, 800Gy of gamma rays). Seeds were abraded with sandpaper to enhance seed germination. LD50 for *P. juliflora* was  $577.875 \pm 67.35$  for gamma-ray and  $18.732 \pm 1.80$  mM for EMS. Seed germination was significantly affected in both species. However, *P. cineraria* proved very sensitive to both mutagens as evident by a very low seed germination thus was not suitable for mutant screening. Nevertheless, *P. juliflora* seed

germination was significantly affected at high doses as compared to medium and low doses. *P. juliflora* seedlings were selected in the field for superior plant height, plant type, stem shape, and diameter as compared to the controlled seedlings. Selected seedlings were further characterized based on calorific value, lignin, cellulose, hemicellulose, moisture, fresh biomass, and dry biomass. Several superior mutants were selected with desirable characteristics for fuelwood, pulpwood, and afforestation. It is concluded that mutation breeding can be used to develop desirable mutants in a short period as compared to conventional breeding. However, future efforts must be directed to refine mutagenic doses for *P. cineraria* to produce a large population for mutant selection and to develop genotypes.

**Keywords:** *Energy Crisis; Pakistan; Deforestation; Climate Change; Prosopis; Calorific Value; Tree Breeding; Genetic Diversity; Mutation Breeding*

## Introduction

Production of energy has become a major issue globally due to a lack of local resources, and low power production, which is causing a major disruption in energy supply. This has been a main challenge as it is negatively impacting the economic, social, and political sectors of Pakistan (Song et al., 2017). Pakistan's energy crisis, caused by increased electricity demand due to population growth, urbanization, and economic expansion, has shrunk the country's GDP (EAW, 2013; Moral-Carcedo and Vicens-Otero, 2005). Therefore, Pakistan is heavily dependent on fossil oil imports, and increasing currency inflation and disruption of supply have further negatively impacted the economic growth of Pakistan. Many industries were forced to move to neighboring countries such as Bangladesh as a result millions lost jobs. Due to power shortages, most industries and households had to rely on biomass for energy generation. Unfortunately, most of the wood biomass is extracted from natural and state plantations illegally or without implementing conservation and management practices. Pakistan is one of the main deforested countries in the world and faces many risks due to climate change. Climate change is increasing the temperature, disrupting water cycles, causing flooding, and prolonged droughts. A recent, flurry of floods in Pakistan is one of the examples of climate-related anomalies in Pakistan. These climate anomalies are having severe and negative impacts on the productivity challenges in major crops threatening food security in Pakistan.

Forests in Pakistan provide vital ecosystem services, including timber, fuel wood, water regulation, and soil protection, among others (Khan and Mehmood, 2003; Mahmood, 2003; Qazi, 1994). However, with only 4.6 million hectares of forest cover and high rates of

deforestation, Pakistan is considered one of the most forest-deficient countries globally (Yaqoob, 2018). Deforestation has caused environmental, floods, health, and financial challenges, including food shortages and high climate risks, making Pakistan one of the top 10 nations facing climate risks in the last two decades (Eckstein et al., 2020). Recent attempts to combat deforestation have failed, leading to extreme weather conditions such as floods, droughts, high temperatures, and heavy snowfalls, exacerbating food shortages and other challenges. Generally, energy is mainly produced from hydro, nuclear, solar, and wind power however the output is not enough to cope with the demand.

Pakistan's energy demand has quadrupled between 2000 and 2020, with an expected doubling by 2030, leading to more oil imports and further economic burden (Shoukat, 2013). Due to the energy shortage and high fuel prices, industries seek cheaper energy sources. In Pakistan, wood biomass is mainly used in the industrial and residential sectors for energy production, and in the pulp and paper industry. Wood energy accounted for 46% of the country's consumption in 1993-94 and typically constitutes nearly 15% of the total primary energy consumption in developing countries (FAO, 1997b; Trossero, 2002). In 2003, Pakistan consumed 43.761 million cubic meters of wood per year, while forest growth was only 14.4 million cubic meters per year, resulting in an annual loss of 29.361 million cubic meters (Pakistan, 2005). Oil may be depleted within 42 years, natural gas in 61 years, and coal in 133 years if the situation persists (Wang and Han, 2011). Pakistan's irrigated and arable lands are mainly used for the production of food crops and fodder. However, there is a vast swathe of land available, that however not suitable for the production of agricultural crops. These lands can be transformed by planting tree species. Therefore, wood from well-managed forests and reforested degraded lands is considered the best alternative to meet the energy crisis.

Drylands cover 41% of the Earth's land surface area (Práválie, 2016), including 80% of Pakistan's land area (Shah et al., 2011). Arid soils have low organic matter, high calcium carbonate content, low nitrogen content, weak profiles, and overutilization resulting in desertification (FAO, 2016; Lal, 2009; Safriel et al., 2006). *Prosopis* species originating from the hyper-arid areas of the world have the potential to survive under these conditions. Moreover, they are most suitable for energy consumption due to their high calorific value. They are also suitable for afforestation due to their high tolerance for drought, waterlogging, and salinity, with enhanced nitrogen to fix nitrogen.

Prosopis is a thorny flowering plant genus in the Fabaceae family, with 45 species distributed across Africa, the Americas, and Asia, including South Asia and Western Asia (Burnham and Johnson, 2004). Four Prosopis species were introduced in Pakistan, including *P. juliflora*, *P. cineraria*, *P. glandulosa*, and *P. farcta*. *P. cineraria* and *P. juliflora* are more widespread in Sindh and Punjab provinces (Essa et al., 2017). *Prosopis juliflora* controls soil erosion, stabilizes dunes, reduces soil salinity, helps in the improvement of soil fertility, and provides fuelwood, construction timber, feed, and food to livestock (Dave and Bhandari 2013; Walter and Armstrong 2014). It is also an excellent source of produce high-quality charcoal (Oduor and Githiomi, 2013). *Prosopis cineraria* is also used as a cattle feed. (Afifi et al., 2018; Yousif, 2012). The pods, bark, flowers, and leaves are used in traditional medicine for the treatment of a variety of health conditions (Pareek et al., 2015). It can also be intercropped with major crops in an agroforestry system (Mahony, 1994).

*Prosopis juliflora* and *P. cineraria* are notorious for having long thorns, multiple stems, crooked stems, and low biomass making it a less preferred species for planting by the farmers. It is therefore important that genotypes with desirable characteristics should be developed. Conventional tree breeding is time-consuming and unsuitable due to long rotational ages, small flower sizes, and polyploidy. On the contrary, mutation breeding, using mutagens such as gamma rays and chemical mutagens such as ethyl methane sulfonate (EMS), offers a good alternative to develop useful mutants (Goel and Behl, 2005; Miller et al., 1984). Several agents that cause mutations are used to induce desirable genetic alterations at high frequencies, including radiation, chemical mutagens, and ionizing agents (Ahloowalia and Maluszynski, 2001). Physical mutagens such as gamma rays have been extremely effective in causing chromosomal aberrations by single or double DNA-strand breaks, or base substitutions. Whereas chemical mutagens like ethyl methane sulfate (EMS), ethyl nitrosourea, dimethyl nitrosamine (DMN), and diethyl sulfate (DES), act basically on nitrogenous base pairs causing different mutations. Over the past 70 years, 2,543 varieties from 175 different plant species, have been developed through mutation breeding worldwide (Chopra, 2005; Maluszynski et al., 2000). Several studies have reported that chemical mutagens are more efficacious than physical mutagens (Rao and Rao, 1983) other experts observed the contrasting behavior (Tara and Dnyansagar, 1980). However, there are very few examples of mutation breeding in tree species therefore both types of mutagens were employed in this study to examine their efficiency on Prosopis species. The main objectives of the study were to determine the sensitivity of Prosopis species to gamma rays and ethyl methane sulfonate mutagens. Secondly, the selection of

genotypes with desirable phenotypic traits such as straight stem form, plant height, high biomass, lignin contents, high calorific value, and thornless genotypes.

## Methodology

To start a tree breeding program, a suitable plus tree must be selected based on its height, diameter, straight bowl, canopy size, crown symmetry, and tolerance to pests and diseases. So, plus trees were identified in various regions of Punjab, Pakistan, with *P. juliflora* being more abundant than *P. cineraria* as *P. cineraria* was observed mostly in wildernesses and graveyards. After identifying the desired plus trees, their seed pods were collected, labeled, and checked for pests and diseases. The seeds were extracted, cleaned, abraded to rupture the hard seed coat, and pre-soaked for 2 hours before being treated with gamma radiation and EMS.

## Mutagenic Treatment

Seeds from both species were treated with physical and chemical mutagen. They were treated with three different doses of both mutagens each treatment was comprised of three replications. Cleaned normal 6000 seeds were treated under three doses of gamma rays 600Gy, 700Gy, and 800Gy and control of 0Gy. Five hundred seeds were treated in each replication. Seeds were at a Cobalt<sup>60</sup> source at the Nuclear Institute for Agriculture & Biology, Faisalabad, Pakistan. The same procedure was adapted for chemical mutagen treatment where a total of 6000 seeds of both species were treated with 15mM, 25mM, and 35mM of EMS and a control with 0 mM concentration. The seeds were pre-soaked in water overnight before being treated with EMS for 8 hours. Treated seeds were kept at room temperature in labeled plastic trays with filter paper above and below the seeds to keep them moist. Water was replaced daily to avoid any fungus development.

## Seed Germination Percentage

The germination percentage for each treatment was recorded after 8 days and then the final recording after 15 days of treatment, as *P. cineraria* seeds started germinating late compared to *P. juliflora*.

$$\text{Seed germination percentage} = \frac{\text{Seeds germinated}}{\text{Total seeds}} \times 100$$

LD50 for both species was estimated by using the LD50 function of the “*HelpersMG*” package (Girondot, 2019) incorporated in the R programming language (R Development Core Team, 2022).

### **Nursery Establishment and Field Plantation**

A nursery was established at Govt. Forest Potted Nursery in Gojra, Punjab. Germinated seedlings were moved to polythene bags and protected from damage by weeding and pest control. A polytunnel was constructed to protect plants from frost damage. The field was prepared according to recommendations, and one-year-old plants were transplanted, watered, and treated with weedicide and pesticide.

### **Data collection**

#### **Phenotypic Data Collection**

Plants were thoroughly observed for mortality rate and phenotypic traits i.e., leaf color changes, plant height, spine size, stem color, stem number, and stem straightness. Three types of phenotypic data were collected i.e., plant height, stem diameter, and number of stems per plant after one year. Plant height was measured with a measuring tape from the ground to the main stem of the plant. Plant stem diameter was measured with a vernier caliper at a height of 4 inches from the ground. The number of stems of each plant was also counted.

#### **Biochemical Analysis**

Those plants that exhibited desirable phenotypic traits were selected for biochemical analysis. Equal-weight wood samples were collected and prepared for biochemical analysis. Seven parameters were determined i.e., cellulose, hemicellulose, lignin, dry matter, moisture content, fiber content, and calorific value. Moisture content was determined by the oven-dry method per BS EN 13183-1 (BS EN 13183-1, 2002). The crude fiber was determined by the method proposed by Bidwell & Walton (1916). The holocellulose (cellulose and hemicellulose) content was determined by the standard method explained by Timell et al. 1959. While lignin content was determined by Klason method (Galiwango, et al., 2018). To determine the calorific value bomb calorimeter method was used following Nelkon and Parker method (1995).

#### **Macropropagation of Selected Mutants**

Desirable mutants were mass propagated through cuttings by using auxin to stimulate root development (Atif, unpublished data).

## Statistical Analysis

Phenotypic and biochemical variation from the control treatment was compared by using Analysis of Variance (ANOVA) by using “*agricolae*” package incorporated in the R-program (R Development Core Team, 2022), and means of parameters with a significant difference were compared with Duncan’s Multiple Range Test (DMRT).

## Results and Discussion

### General observation

Prosopis species were introduced to Pakistan for dune stabilization and income generation for the poor farmers having marginal lands. *P. juliflora* and *P. cineraria* are the most common Prosopis species, with *P. juliflora* commonly found on roadsides, wastelands, graveyards, and government rangelands. Its sweet pods are eaten by animals, and they help in seed dispersal and germination. *P. cineraria* has small thorns and ash-grey bark and is more common in Southern Punjab. One unique feature was observed in *P. juliflora* that it typically has 3-8 stems per plant, but in some cases, it has a single stem. The seed collection for *P. cineraria* and *P. juliflora* was a challenging task because *P. cineraria* showed a scattered low population while *P. juliflora* trees possess long thorns making it difficult to collect seeds. Vigorous, disease, and pest-free, and preferably single-stemmed trees with good canopies were selected for seed collection. *P. juliflora* was easy to locate, but finding a plus tree was difficult because of its crooked and multi-stem type. *P. cineraria* trees were hard to find, but those found were usually single-stemmed and vigorous.

### Germination Percentage

Table 1 shows the germination percentage of *P. juliflora* and *P. cineraria* respectively. Gamma radiation doses decreased germination percentage as compared to the control in both species, with the most significant reduction observed at 800Gy. Similarly, high chemical mutagen treatment had a negative effect on seed germination as compared to the control treatment, and the lowest germination was observed at 35mM. The results suggested that high levels of mutagens had a negative impact on seed germination regardless of the species.

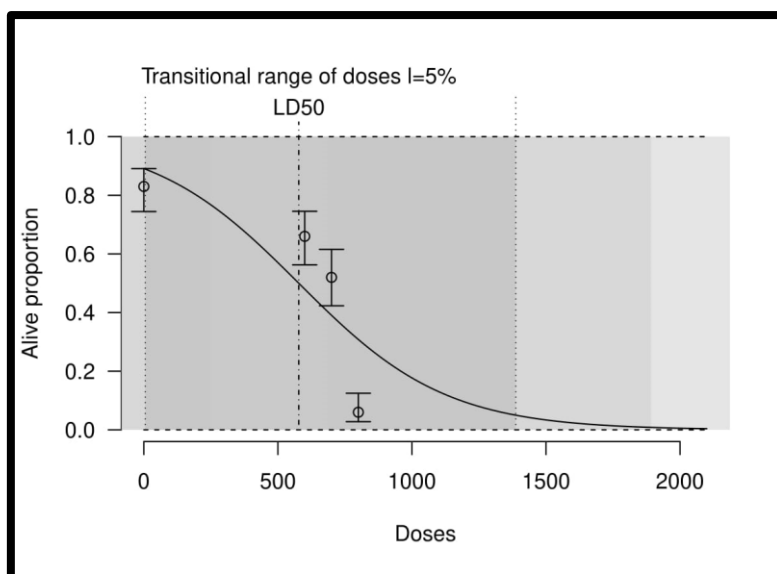
**Table. 1.** Comparison of germination percentages of *P. juliflora* and *P. cineraria*

Treatment	Germination Percentage %	
	<i>P. juliflora</i>	<i>P. cineraria</i>
<b>Physical Mutagen</b>		
Control	83.00%	34.70%
600Gy	66.00%	31.02%
700Gy	52.00%	19.50%
800Gy	5.73%	8.80%
<b>Chemical Mutagen</b>		
15mM	57.00%	39.10%
25mM	39.00%	18.00%
35mM	19.00%	16.40%

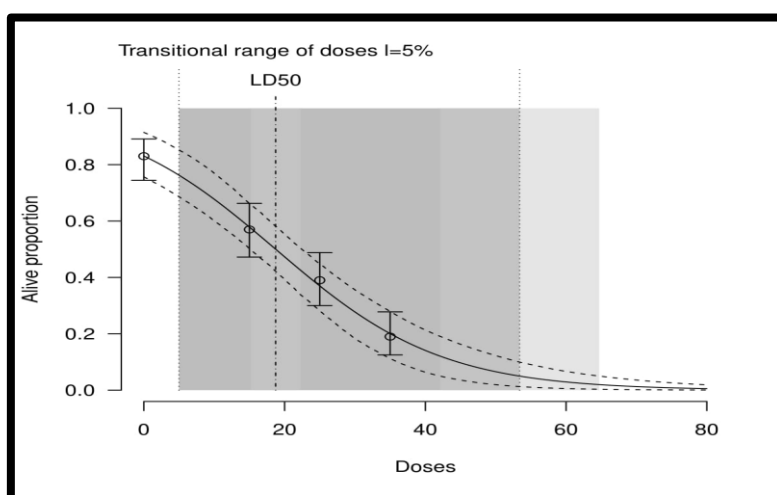
**Median Lethal Dose (LD50)**

LD50 was determined in both species based on the relationship between gamma-ray doses and germination percentage. For *P. juliflora* seeds treated under gamma irradiation, the LD50 based on seed viability was  $577.875 \pm 67.353$  (Fig. 1) and LD50 for EMS treatment was calculated to be  $18.732 \pm 1.803$  mM (Fig. 2).





**Figure 1.** LD50 of *P. juliflora* seeds from gamma irradiation treatment



**Figure 2.** LD50 of *P. juliflora* seeds treated with ethyl methane sulfonate (EMS) treatment.

### Plant Mortality

Higher doses of mutagens resulted in higher mortality rates, with the highest at 62.79% in 800Gy gamma radiation. EMS doses showed an upward trend in mortality rate, with the highest at 57.54% from 35mM. *P. cineraria* had the highest mortality rate with all treatments showing above 90% mortality. Plants showed stunted growth, thin stems, and wilting. Among the observed plants, the control group exhibited the least impact, displaying a mortality rate of 90.98%. Conversely, the plants subjected to 800Gy gamma irradiation demonstrated the

highest level of susceptibility, resulting in a mortality rate of 97.08%. Due to poor seed germination and a high mortality rate in *P. cineraria*, further studies were carried out on *P. juliflora*.

### Phenotypic Data

Table 2 displays the findings derived from an investigation conducted to analyze the growth patterns of *P. juliflora* when subjected to various treatment conditions. The mutagenic treatment had a significant effect (F-Value = 380.8,  $p < 0.001$ ) on plant height. The control group had the tallest plants (48.28 cm), while the 25 and 35 groups had the shortest plants 24.91 and 24.95 cm, respectively. The number of stems did not show a significant difference between treatments. For diameter, the treatment also had a significant effect (F-Value = 24.52,  $p < 0.001$ ). The control group had the largest diameter (0.615 cm), while the 800Gy and 600Gy groups had the smallest diameter 0.521 and 0.537 cm, respectively. The 15mM and 35mM had intermediate diameters (0.584, 0.575, and 0.545 cm, respectively). Although the 700Gy treatment group was not the highest significant in terms of plant height, a plant from this group showed the highest stem height of 71cm among all groups including the control. Similar plants also possessed the highest diameter value of 1.2cm (Fig. 3).

Overall, the results suggest that high gamma radiation and EMS doses had a negative effect both on height and diameter. However, some plants from different mutagens treatment groups showed superior growth in height and stem diameter. The number of stems per plant was not significantly affected by either treatment or concentration. The control group showed the best growth performance in terms of plant height and diameter.

**Table. 2.** Comparison of phenotypic parameters of *P. juliflora* through Analysis of Variance (ANOVA).

Parameter	SOV	Df	SS	MS	F Value	Duncan's Comparison Test		Mean
Plant Height	Treatment	6	139093	23182	380.8***	Control	48.28a	
						600Gy	34.05b	
						15mM	31.01c	
						700Gy	29.40c	
	Residuals	3221	196086	61		800Gy	26.40d	
						35mM	24.95e	
						25mM	24.91e	
Number of Stems	Treatment	6	2	0.03074	0.153 ns			
	Residuals	3221	6474	2.0099				
Diameter	Treatment	6	2.41	0.4021	24.52***	Control	0.615a	
						700Gy	0.584b	
						15mM	0.575b	
						35mM	0.545c	
	Residuals	3221	52.81	0.0164		25mM	0.538c	
						600Gy	0.537c	
						800Gy	0.521c	

\* p≤0.05

\*\* p≤0.01

\*\*\* p≤0.001



**Figure 3.** Growth comparison of A) *P. juliflora* plant treated with 700Gy of gamma radiation B) Control *P. juliflora* plant.

### **Gamma Irradiation and Biochemical Parameters**

This table (Table 3) presents the results of an analysis of variance (ANOVA) for biochemical parameters of *P. juliflora* plants treated under gamma radiation. From the table, we can see that there are significant differences between the treatment groups for some of the parameters. For example, the dry matter content and moisture content show significant differences between the treatment groups ( $p < 0.05$ ) and control, while the crude fiber, cellulose content, and hemicellulose content do not show significant differences ( $p > 0.05$ ). The lignin content and calorific value also show significant differences between the treatment groups and control ( $p < 0.05$ ).

**Table 3.** Comparison of biochemical parameters of *P. juliflora* plants treated under gamma radiation through Analysis of Variance (ANOVA).

Parameter	SOV	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Dry matter content	Treatment	3	3.234	1.0782	5.433	0.00905**
	Residuals	16	3.175	0.1984		
Moisture content	Treatment	3	3.234	1.0782	5.433	0.00905**
	Residuals	16	3.175	0.1984		
Crude Fiber	Treatment	3	1.88	0.6267	0.241	0.867 <sup>ns</sup>
	Residuals	16	41.62	2.6013		
Cellulose Content	Treatment	3	8.87	2.956	0.504	0.685 <sup>ns</sup>
	Residuals	16	93.83	5.865		
Hemicellulose Content	Treatment	3	12.25	4.084	0.587	0.632 <sup>ns</sup>
	Residuals	16	111.29	6.955		
Lignin Content	Treatment	3	26.19	8.73	3.444	0.042*
	Residuals	16	40.56	2.535		
Calorific Value	Treatment	3	26820	8940	3.374	0.0446*
	Residuals	16	42400	2650		

### EMS Treatment and Biochemical Parameters

Table 4 showcases the outcomes obtained from an analysis of variance (ANOVA) performed to assess the biochemical parameters of *P. juliflora* plants subjected to EMS treatment. Based on the data presented in the table, it is evident that no statistically significant differences exist among the treatment groups for any of the parameters examined ( $p > 0.05$ ). The dry matter content, moisture content, crude fiber, cellulose content, hemicellulose content, lignin content, and calorific value do not show significant differences between the treatment groups. Therefore, the EMS treatment does not seem to have a significant effect on the biochemical parameters of *P. juliflora* plants.

**Table 4.** Comparison of biochemical parameters of *P. juliflora* plants treated with EMS through Analysis of Variance (ANOVA).

Parameter	SOV	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Dry matter content	Treatment	3	1.23	0.41	1.829	0.174 <sup>ns</sup>
	Residuals	20	4.483	0.2242		
Moisture content	Treatment	3	1.23	0.41	1.829	0.174 <sup>ns</sup>
	Residuals	20	4.483	0.2242		
Crude Fiber	Treatment	3	12.04	4.014	1.469	0.253 <sup>ns</sup>
	Residuals	20	54.64	2.732		
Cellulose Content	Treatment	3	10.86	3.621	0.659	0.587 <sup>ns</sup>
	Residuals	20	109.94	5.497		
Hemicellulose Content	Treatment	3	28.46	9.486	1.197	0.336 <sup>ns</sup>
	Residuals	20	158.54	7.927		
Lignin Content	Treatment	3	14.83	4.944	1.825	0.175 <sup>ns</sup>
	Residuals	20	54.18	2.709		
Calorific Value	Treatment	3	6553	2184	0.873	0.472 <sup>ns</sup>
	Residuals	20	50071	2504		

### Selection of Superior Genotypes

Table 5 shows selected mutant plants of *P. juliflora* with different phenotypic and biochemical parameters. Several useful mutants were selected exhibiting desirable traits such as plant height, diameter, and low and high lignin contents. Plant a379, from 600Gy gamma rays' treatment, shows the highest number of stems per plant, which is 8. A mutant b127, from 700Gy treatment, showed a single stem and the highest lignin and calorific value. Another plant b253 from the same group resulted in the highest value of dry matter and cellulose. Mutant y203, from 25mM EMS treatment, showed 0.4cm diameter but the same plant gave 5000kcal/kg calorific value and 6 stems. A mutant from 35mM EMS treatment showed 7 stems-per-plant, it also possessed the lowest lignin content of 12.6% and the lowest calorific value was 4790kcal/kg, making it a suitable choice for the paper industry.

**Table 5.** Phenotypic and Biochemical data of selected mutant *P. juliflora* plants

Tag #	Treatment	Phenotypic Data			Biochemical Data						Calorific Value
		Height (cm)	No. of Stems	Diameter (cm)	% Dry matter	% Moisture content	% Crude Fiber	% Cellulose	% Hemicellulose	% Lignin	Calorific Value (Kcal/kg)
a379	600Gy	46	8	0.7	95.5	4.5	66	46	23.2	14	4860
b127	700Gy	71	1	1.2	97	3	65.3	41.7	29	19.3	5040
b253	700Gy	25	7	0.5	97	3	64.9	47	24.3	18.5	5000
y203	25mM	31	6	0.4	95.8	4.2	66	38.9	25.9	18.8	5000
z75	35mM	29	7	0.4	96.6	3.4	66.5	43.5	30.5	12.6	4790

## Discussion

Prosopis was introduced in Sindh, Pakistan for afforestation but became an invasive weed due to its deep roots and drought tolerance. It is still preferred for afforestation in low-water availability and high-temperature regions. Its uses include fuelwood, charcoal, and fodder. Its drawbacks are its short height, low biomass, stem diameter, invading nature, long thorns, and multi-stem characteristics. So, after an in-depth and detailed observation of its uses among local communities and afforestation potential, the main purpose of conducting this study was the selection of mutants with desirable features such as a single stem, large diameter, high and low calorific value, and lignin contents, and thorn lessness. The evaluation of the LD50 of a substance or mutagen is necessary to identify any detrimental effects resulting from deliberate or accidental short-term exposure (Clemedson et al., 2000). So, LD50% for both mutagens was determined. LD50% of gamma rays for Prosopis were 750Gy as determined by Asif et al. (2020) and LD50% of EMS was studied in this experiment, which was 25mM.

Physical and chemical mutagens negatively affected seed germination due to their destructive features. Low gamma irradiation dosages resulted in high seed germination percentages, but this decreased at higher doses (700Gy and 800Gy). *P. cineraria* had the lowest germination percentage even among the control treatments. These results are like the findings of many previous investigators who described that higher doses of gamma radiation decrease seed germination percentage in different crop and tree plants. For example, a very high dose of 50Gy of gamma rays on barley seeds by Wang et al. (2017) suppressed seed germination percentage. Similarly, a 400Gy dose in fenugreek by Hanafay and Akladius (2018), a 200Gy dose in *Zea mays* by Marcus et al. (2013), and a 300Gy dose of gamma rays by Norfadzrin et al. (2007) in tomato seeds resulted in lower germination percentage as compared to lower doses. Goel and Behl (2005) also documented a corresponding inverse relationship in the seed germination percentage and gamma radiation dose in seeds of two semi-arid tree species, *Prosopis juliflora*, and *Acacia nilotica*.

EMS treatment showed the same trend that higher doses produced low germination. A similar result was obtained by Laskar et al. (2018) when they treated tomato seeds with EMS and recorded low germination in M1 at higher concentrations. Some more examples of studies with corresponding results of low germination from higher EMS concentrations are Ambli and Mullainathan (2014) in pearl millet, Anbarasan et al. (2013) in sesame, Ariramana et al. (2014) in



pigeon pea, Gnanamurthy et al. (2011) in maize, Satpute and Fultambkar (2012) in soybean, Talebi et al. (2012) in rice and Talebi et al. (2012) in cluster bean. This decrease in germination against higher doses of gamma rays can be due to subsequent mitotic abnormalities, seed dormancy, moisture content, seed maturity, seed tissue damage, and chromosome retardation (Al-Safadi and Simon 1990; Datta, 2009). Similarly, Khan and Goyal (2009) stated that this low germination can be due to altered enzyme activity or damage to cell components.

Plant mortality increased with increasing doses of EMS and gamma radiation in *P. juliflora*. Control group mortality was far lower 800Gy of gamma radiation and 35Mm of EMS. *P. cineraria* had high mortality rates (above 90% in all mutagen treatments). Mortality started at 800Gy gamma irradiation and 35mM EMS, moving down towards lower doses, with control plants being the last to die. These results are in accordance with many studies carried out previously as a treatment of high doses in *Withania* by Das et al. (2010) decreased the plant survival rate. Soybeans showed a high mortality rate in M1 and M2 after treatment with gamma radiation and mortality increased with dose (Mehetre and Kshirsagar, 2022). Goel and Behl (2005) also documented the direct relation of increasing doses with mortality rate. Similarly, cowpeas by Girija and Dhanavel (2009) and horse gram by Kulkarni (2011), chickpeas by Barshile (2006) were treated with different EMS concentrations and all these researchers recorded low plant survival at higher doses of EMS. This reduction in plant survival can be due to severe damage to the plant cells, as reported by Sikora et al. (2011) that point mutation at random sites is caused by EMS which retards many processes of protein synthesis.

Gamma rays and EMS concentration affected the stem heights of *P. juliflora* plants. Lower doses of both mutagens resulted in greater mean stem heights than higher doses. Physical mutagen at a low dose of 600Gy resulted in more mean height than a low dose of 15mM EMS. In general, higher doses resulted in lower stem height. These findings are in accordance with previous studies. Gamma radiation treatment by Irfaq and Nawab (2001) on three wheat varieties resulted in a decrease in the survival rate and a delay in germination. Plant height also decreased with a higher dose of gamma rays. Similarly, Kalia et al. (2001) in bread wheat, Gonzales (2007) in orchids, and Yaping, (1996) in chickpeas reported the same behavior of plant height reduction at higher doses of EMS. Shah et al. (2008) stated that some processes like changes in ascorbic acid content, biochemical and physiological disturbances, and destruction of auxin, can cause inhibition of plant

development and germination rate. In consensus with our findings on plant height, Behera et al. (2012) also reported some morphological effects of EMS treatment like leaf size and plant height in *Asteracantha longifolia*. Many previous studies suggested that low plant height at higher dosages of mutagens can be due to the reason of abnormal production of different signaling hormones like indole acetic acid, gibberellin, or strigolactones (Bennet et al., 2014; Bishop, 2003, Kwon and Choe, 2005; Liu et al., 2010 and Sazuka et al., 2009).

Physical mutagen significantly affected the *P. juliflora* plant stem diameter positively and the diameter increased from lower to higher doses of gamma radiation. On the other hand, EMS or chemical mutagen showed a negative or declining trend in mean stem diameters. Higher doses of EMS produced lower stem diameters as the lowest dose of 15mM gave the highest mean stem diameter. Similar results were also obtained by Wi et al. (2007) as larger stem diameters were noticed from higher doses of 8krad of gamma radiation. Goel and Behl (2005) also observed an increase in diameter at higher doses of gamma rays. Some other researchers like Fu et al. (1999) stated the opposite results as diameter decreased for higher doses of gamma radiation. Chaomei and Yanlin (1993) and Melki and Marouani (2010) also reported a decrease in plant vigor at higher doses. This increase in stem diameter is possibly because gamma rays negatively affected the stem height from lower to higher doses which forced lateral meristematic growth which resultantly increased the stem diameter. Behera et al. (2012) reported the detrimental effects of EMS on plant vigor at higher doses which were like our results of EMS effect on plant diameter. Ibrahim (2008) tested EMS effects on strawberries and reported the same behavior of higher doses (reduction in plant vigor).

The number of stems per plant was not affected negatively or positively, neither by gamma radiation nor by ethyl methanesulphonate, as all the groups showed nearly the same mean value for the number of stems per plant (3.93 to 4.06) in comparison to the control group's value of 4.08. Previous studies by different researchers reported three types of contrary effects (no effect, increasing, and decreasing) on stems per plant by increasing the dose of gamma irradiation. Tatiana et al. (2019) reported no significant variation in stem number in *Calendula officinalis* from different treatments of chemical mutagens. Kaul and Bradu (1972) found an increasing number of stems or branches per plant after irradiation treatment of seeds of *Atropa belladonna*. Similarly, Alba et al. (2004) in *Chamomilla recutita*, and Youssef et al. (2000) in geranium obtained the same

results. In the case of chemical mutagenesis, Jabeen and Mirza (2002), and Sri Devi and Mullainathan (2011) reported the same response of an increasing number of branches or stems at higher doses of EMS. In contrast to these findings, Marizka (2010) in soybean, and Shukla and Datta (1993) in Chrysanthemum reported that mutagenic dose and the number of stems per plant are inversely proportional. To put it simply, increasing the gamma radiation dose decreased the number of branches or stems. Stem numbers did not show significant variation, indicating the potential for generating mutants with desirable height and diameter characteristics. Therefore, the same mutagenic dosage can be used to develop mutant *P. juliflora* plants with improved height and diameter without altering stem numbers. Although, there was no significant difference among the mean number of stems of different treatment groups a plant from 600Gy gave 8 stems per plant. This trait could be useful in arid area afforestation due to their wider canopy and greater biomass production.

The dry matter content of *P. juliflora* plants increased gradually from 0Gy to 700Gy, then declined to 800Gy. EMS-treated groups exhibited increased dry matter content mean values for all dosages. However, the difference among chemical mutagen doses was insignificant. These findings are consistent with previous research, which demonstrated that radiation improves dry matter content until a specific dose, after which it declines at higher lethal doses. Sarker et al. (2014) reported a decrease in total dry matter content in soybeans at higher doses of 300Gy, 400Gy, and 500Gy of gamma rays as compared to control and lower doses of 200Gy. Some other investigations by Bamidele and Akanbi (2013), Linda et al. (2020), and Rao et al. (1994) did not report any significant effect on dry matter percentages from different levels of gamma radiation dosages. Mutants with increased dry matter content are valuable in this regard because dry matter content is a crucial indicator of nutrient availability and energy content in wood. It is also essential for biomass production, carbon sequestration, and pulp production. Plants with higher dry matter content are preferred in the paper industry, as the water content is considered waste in pulp production.

Gamma radiation did not significantly affect the moisture content percentage in *P. juliflora* plants. The lowest dose of 600Gy slightly decreased the moisture content as compared to the control group. Then 700Gy gave a lower moisture content as compared to the last dose of 600Gy but the 800Gy group gave the highest mean value of 4.25% (more than the control). In the case of ethyl

methanesulfonate, all dosages generated a mean value between 3.53% (35mM) to 4.11% (control) so the overall effect was non-significant for both types of mutagens. These results are in accordance with the findings of Bamidele and Akanbi (2013), Linda et al. (2020), and Rao et al. (1994) as they obtained no significant effect of gamma irradiation on moisture content in *Cajanus cajan*. However, some other researchers Arthur and Wiendl, (2000) and Premi Devi et al. (2018) reported contrary results of the significant reduction in moisture content at higher doses of gamma radiation. The moisture content of wood is very important from the perspective of the paper industry as well as for plantation projects in drier conditions of arid or semi-arid areas. Plants possessing a higher percentage of water content due to their genetic mutations would prove to be advantageous for cultivation in regions experiencing water scarcity. This is because they possess the capability to retain a larger quantity of water during the rainy season. Plants with less moisture content would be a good choice for pulp and paper manufacturing and timber purposes. Because high water content causes timber wood to shrink and then expand.

Crude fiber percentage was not significantly affected by gamma radiation. However, the highest dose of 800Gy slightly decreased the fiber content. So, in general, all the treatments produced almost the same mean fiber percentage. EMS dose of 35mM gave the highest mean fiber value and the lowest dose of 15mM produced the lowest mean value among all the groups. As all mutagenic treatments failed to induce significant differences, the crude fiber remained unaffected by the treatments. Findings by Ihsanullah et al. (2005) and Mohammadzai et al. (2010) were in support of our results as they observed no significant change in crude fiber content in dates, both with lower and higher doses of gamma radiation. In contrast to these results, some researchers like Mohammadzai et al. (2010) and Abdelwhab et al. (2009) reported a negative effect on crude fiber percentages at higher doses when they treated beans and palm dates under gamma exposure, respectively. There are no or very few studies on crude fiber percentage from EMS treatments. However, Al-Rawi and Kohel (1970) and Witt et al. (2018b) reported the positive effect of EMS on different traits of fiber i.e., improvement of fiber quality, length of fiber, strength, and elongation of fiber in cotton.

Gamma radiation and ethyl methanesulfonate did not affect the holocellulose content significantly. However high doses of both mutagens reduced the holocellulose content as compared to lower dosages. Some previous studies, Ajila et al. (2007), Emaga et al. (2008), and Orozco et al. (2012)

also showed degradation of holocellulose at higher doses of gamma rays. Similar results of reduced cellulose were obtained by Purente et al. (2020) from different higher concentrations of ethyl methanesulfonate. As in our study cellulose content of *P. juliflora* in both types of mutagenic treatments was higher than the control group at lower doses so 600Gy dose of gamma radiation and 15mM dose of EMS can be further used to develop a mutant with higher cellulose content which can be used for pulp production or paper making. According to Molin and Teder (2002), a high ratio of cellulose to hemicellulose increases folding endurance and fracture toughness. Hemicellulose on the other hand can be used for sorbitol (pharmaceutical sweetener) production by reduction process and alcohol production by fermentation (Gírio et al., 2010).

Lignin percentage was affected significantly by gamma radiation and a moderate dose of 700Gy gave the highest mean value of 17.88%. A non-significant effect was observed by EMS treatment, however, the highest dose of 35mM decreased the lignin content as compared to the control and other groups. Results from physical mutagenesis are corroborated by the findings of Chin et al. (2017) where lignin content decreased at higher doses of gamma radiation. Similarly, the results of Purente et al. (2020) support our findings that lignin decreased by increasing EMS dosages. In the paper industry, the removal of lignin from pulp is not specifically targeted for application purposes but to enhance the paper quality (Cotana et al., 2014) because lignin gives yellowish color to paper. So, lignin is removed from wood pulp by Kraft pulping, soda pulping, or sulfite pulping methods (Haq et al., 2020). But these methods are harsh, expensive, and time-consuming so mutants with low lignin content can be used for pulp production. On the other hand, lignin is positively and significantly correlated with calorific value. This mutant with high lignin content can be used as fuelwood as reported by Beauchet et al. (2012). Lignin can be yielded (from pulp) from mutants with high lignin percentage which can be used as a biofuel, bioplastic, biofertilizer, value-added chemicals, food additives, animal feed, textile, personal care product, adhesives, and lubricants (Cho et al., 2018; de Wild et al., 2017). Moreover, improved genotypes with high biomass yield and better phenotypic traits would be accepted and preferred by the local community.

Changes in calorific values in different wood species can be accredited to the biochemical composition of the wood. Some extractive contents and most importantly lignin percentage show a notable influence on calorific value. As stated by Kaltschmitt and Streicher (2009), the net heat

of combustion from lignin is 27.0 MJ/kg while cellulose and hemicelluloses have 17.3 and 16.2 MJ/kg net heat of combustion, respectively. Similarly, Fuwape (1989) obtained 25.4 and 19.7 MJ/kg gross-heat combustion values for lignin and cellulose in *Gmelina arborea*. Therefore, the higher the lignin content, the higher will be the calorific value (Demirbas, 2001; Kataki and Konwer, 2001). Moreover, Krauss et al. (2011) and Wagenführ (2007) corroborated these results by getting a higher net heat of combustion value of 19.1 MJ/kg from Santos rosewood and a lower value of 16.6 MJ/kg from Birchwood because Santos rosewood has approximately 31.2 % lignin while Birchwood possesses lignin content between 19.3 and 27.4 percent. According to Demirbas (2001), the higher heating value (HHV) of any lignocellulosic fuel from any plant species is in fact a function of its percentage of lignin. Howard (1973) reported that cellulose and hemicelluloses possess a higher heating value of 18.60 kJ/g, on the other hand, lignin alone has about 23.26 to 25.58 kJ/g HHV.

For mutant plant selection, this study used phenotypic traits like height, diameter, and number of stems. Straight and tall single-stemmed plants were selected for timber production, while lower-height, higher-diameter plants were chosen for biomass. Multi-stemmed plants were picked for afforestation and reforestation projects and were found to have more green foliage, suitable as forage in areas with shortages. Selected mutants underwent biochemical analysis for desirable traits. High-lignin plants were good for energy production, and low-lignin for pulp and paper. Plants with high holocellulose levels were ideal for biomass, and high moisture for arid lands. Mutants with high calorific values were suitable for charcoal production or fuel. These traits have the potential for economic and environmental benefits in various sectors.

In this study gamma radiation proved to be more effective as compared to EMS treatment on seeds of both *Prosopis* species. The following reasons provide insights into the increased effectiveness of gamma radiation in comparison to chemical mutagens. Gamma radiation can cause more extensive DNA damage and a higher incidence of mutations because it has high energy and can enter the target tissue deeply. Chemical mutagens, like EMS, on the other hand, frequently have limited ability to enter the target cells or tissues. For example, a study on tomatoes by Shirasawa et al. (2016) stated that gamma rays caused a higher percentage of deletions and insertions in the tomato genome as compared to EMS.

As mentioned earlier, gamma radiation induces a variety of mutations, including translocations, large deletions, and chromosomal aberrations which result in more unique changes in phenotype or gene expression. On the other hand, chemical mutagens mostly induce point mutations, which result in less effect on gene function. Xiao et al. (2019) stated that EMS usually induces nucleotide modifications causing mispairing of C with T, which leads to the transition of CG to TA. Mutagens can differ in their level of effectiveness for different species. This is because of transformations in repair mechanisms of genetic makeup. For instance, the Sofu cultivar of *Triticum durum* showed a higher frequency of mutation when treated with gamma irradiation as compared to EMS treatment, while another cultivar Gediz-75 from *Triticum durum* displayed a higher mutation rate when treated with EMS (Sakin and Sencar, 2002).

Different cells or tissues of a plant species can have different sensitivities to different mutagenic agents. For instance, some tissues may be more sensitive to chemical mutagens, while some other cells or tissues may be more susceptible to gamma rays. Effectiveness may decrease or increase when a combination of different mutagens is used. A study on tomatoes showcased the highest effectiveness of gamma radiation followed by sole EMS and a combination of EMS and gamma radiation treatments (Sikder et al., 2013). Multiple copies of each chromosome exist in polyploid plants, which can enhance the risk of genetic alterations brought on by mutations and provide genetic redundancy. As a physical mutagen, gamma radiation can alter chromosomes in a variety of ways, including deletion, translocation, inversion, and duplication, which increases genetic variability in polyploid plants. EMS, in contrast, mostly causes point mutations, which are less likely to result in significant genetic changes in polyploid plants. Quantitative traits are controlled by multiple genes so mutations like point mutations by EMS affecting just one or few genes may not develop a significant alteration in plant phenotype especially in polyploids like *P. juliflora*. Nevertheless, gamma radiation with its broad spectrum of effects and power can cause transformations in multiple genes at the same time, resulting in noteworthy mutations in quantitative characters. While EMS induces just point mutations, resulting in less substantial vagaries in phenotype.

The composition of alleles may offer a different response of plant species to mutagens. While gamma rays induce mutations in dominant as well as recessive alleles, EMS primarily causes mutations in recessive alleles making EMS less effective in this case. Therefore, gamma rays

generate a broader range of mutations enhancing the possibility of obtaining beneficial traits. Finally, gene action can also affect the mutagenic response of plant species. Gamma radiation can induce qualitative along with quantitative mutations in gene expression, but EMS mutations mainly affect negligible quantitative alterations in gene expression. Therefore, gamma radiation produces diverse and extensive mutations in gene expression, which may result in increased probabilities of getting advantageous characteristics.

## Conclusions

*P. juliflora* has a narrow genetic base and its importance can be enhanced by using breeding techniques but due to the long rotation age of trees, it is a very difficult and time-consuming task to use conventional breeding techniques. So, mutation breeding and biotechnology techniques are a good way to obtain good mutants with desirable phenotypic characteristics. Mutation breeding has been successfully used in crops to produce new varieties in a short period. Mutation breeding has only recently been used in tree species. Both physical and chemical mutagens are used in a mutation breeding program. However, physical mutagens are commonly used due to ease and safety as compared to chemical mutagens. Gamma rays are preferred as a physical mutagen whereas EMS is commonly used as a chemical mutagen. Both mutagens were used in this study due to their different modes of action on the genome. Physical mutagen proved more effective in inducing genetic diversity as compared to chemical mutagen. This could be due to the mode of action of these mutagens and the hard seed coat of the species. *P. juliflora* is found commonly and widely thriving in nature as compared to *P. cineraria*. A restricted distribution in the wild could be attributed to a very low seed germination rate that was observed in *P. cineraria* as compared to *P. juliflora*. As a result of low seed germination and low population size, further studies were conducted on *P. juliflora*. A wide range of responses was observed when *P. juliflora* seeds were treated with mutagens, such as low seed germination, high mortality, stunted growth, plant height, stem diameter, and difference in biochemical composition of the mutants. Several mutants with desirable traits such as tall and short, different stem diameters, low to high cellulose content, variable lignin content, and calorific value were successfully selected. These mutants prove superior to the control plants. This showed that mutation breeding can be used in tree species to develop desirable genotypes in a short period. Selected mutants were asexually multiplied for field planting and evaluation.



## References

1. Abdelwhab, N.M., Nour, A.M. and Fageer, A.S.M., 2009. The nutritive and functional properties of dry bean (*Phaseolus vulgaris*) as affected by gamma irradiation. *Pak. J. Nutr*, 8(11), pp.1739-1742.
2. Afifi, H.S.A. and Al-rub, I.A., 2018. *Prosopis cineraria* as an unconventional legumes, nutrition and health benefits. In *Legume seed nutraceutical research*. IntechOpen.
3. Ahloowalia, B.S. and Maluszynski, M., 2001. Induced mutations—A new paradigm in plant breeding. *Euphytica*, 118, pp.167-173. Ajila, C.M., S.G. Bhat and U.J.S. Prasada. 2007. Valuable components of raw and ripe peels from two Indian mango varieties. *Food Chem.* 2007, 102, 1006–1011.
4. Al-Qurainy, F. and Khan, S., 2009. Mutagenic effects of sodium azide and its application in crop improvement. *World Applied Sciences Journal*, 6(12), pp.1589-1601.
5. Al-Rawi, K.M. and Kohel, R.J., 1970. Gene Action in the Inheritance of Fiber Properties in Intervarietal Diallel Crosses of Upland Cotton, *Gossypium hirsutum L.* 1. *Crop Science*, 10(1), pp.82-85.
6. Al-Safadi, B. and Simon, P.W., 1990. The effects of gamma irradiation on the growth and cytology of carrot (*Daucus carota L.*) tissue culture. *Environmental and experimental botany*, 30(3), pp.361-371.
7. Anbarasan, K., Sivalingam, D., Rajendran, R., Anbazhagan, M. and Chidambaram, A., 2013. Studies on the mutagenic effect of EMS on seed germination and seedling characters of Sesame (*Sesamum indicum L.*) Var. T MV3. *International Journal of Research in Biological Sciences*, 3(1), pp.68-70.
8. Ariraman, M., Gnanamurthy, S., Dhanavel, D., Bharathi, T. and Murugan, S., 2014. Mutagenic effect on seed germination, seedling growth and seedling survival of Pigeon pea (*Cajanus cajan (L.) Millsp.*). *International Letters of Natural Sciences*, 16.
9. Arthur, V. and Wiendl, F.M., 2000. Gamma irradiation of starfruit to increase shelf life. *Revista de Agricultura (Piracicaba)*, 75(3), pp.425-429.
10. Arulbalachandran, D., Mullainathan, L. and Velu, S., 2009. Screening of mutants in black gram (*Vigna mungo (L.) Hepper*) with effect of DES and COH in M2 generation. *J. Phytol*, 1, pp.213-218.
11. Asif, J.M., Ali, A., Mazhar, M.Z., Tanvir, A., Zia, B., Anmbreen, I., Anjum, M.Z. and Mahr, M.S., 2020. Effect of different pre-treatments on seed germination of *Prosopis juliflora* and *Dalbergia sissoo*: a step towards mutation breeding. *Journal of forest science*, 66(2), pp.80-87.
12. Bamidele, O.P. and Akanbi, C.T., 2013. Effect of gamma irradiation on physicochemical properties of stored pigeon pea (*Cajanus cajan*) flour. *Food science & nutrition*, 1(5), pp.377-383.
13. Barshile, J.D., 2006. Studies on effect of mutagenesis employing EMS, SA and GR in Chickpea (*Cicer arietinum L.*) (Doctoral dissertation, Ph. D. Thesis).
14. Beauchet, R., Monteil-Rivera, F. and Lavoie, J.M., 2012. Conversion of lignin to aromatic-based chemicals (L-chems) and biofuels (L-fuels). *Bioresource technology*, 121, pp.328-334.
15. Behera, M., Panigrahi, J., Mishra, R.R. and Rath, S.P., 2012. Analysis of EMS induced in vitro mutants of *Asteracantha longifolia (L.) Nees* using RAPD markers.

16. Bennett, T. and Leyser, O., 2014. Strigolactone signalling: standing on the shoulders of DWARFs. *Current opinion in plant biology*, 22, pp.7-13.
17. Bishop, G.J., 2003. Brassinosteroid mutants of crops. *Journal of Plant Growth Regulation*, 22, pp.325-335.
18. BS EN 13183-1. 2002. The moisture content of a piece of sawn timber. BS EN - British Standard Institute: London, UK, 2002.
19. Burnham, R.J. and Johnson, K.R., 2004. South American palaeobotany and the origins of neotropical rainforests. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 359(1450), pp.1595-1610.
20. Chaomei, Z. and Yanlin, M., 1993. Radiation-induced changes in enzymes of wheat during seed germination and seedling growth. *Henong Xuebao (Acta Agriculturae Nucleatae Sinica);(China)*, 7(2).
21. Chin, S.X., Chia, C.H., Zakaria, S., Ahmad, S.H. and Tasirin, S.M., 2017. Combination of gamma irradiation and sodium carbonate pretreatment on oil palm empty fruit bunch (EFB) for high acidic hydrolysis yield. *Sains Malaysiana*, 46(1), pp.167-173.
22. Cho, M., Karaaslan, M., Wang, H. and Renneckar, S., 2018. Greener transformation of lignin into ultralight multifunctional materials. *Journal of Materials Chemistry A*, 6(42), pp.20973-20981.
23. Clemedson, C., Barile, F.A., Chesne, C., Cottin, M., Curren, R., Ekwall, B., Ferro, M., Gomez-Lechon, M.J., Imai, K., Janus, J. and Kemp, R.B., 2000. MEIC evaluation of acute systemic toxicity. Part VII. Prediction of human toxicity by results from testing of the first 30 reference chemicals with 27 further in vitro assays. *ATLA*, 28(1), pp.161-200.
24. Cotana, F., Cavalaglio, G., Nicolini, A., Gelosia, M., Coccia, V., Petrozzi, A. and Brinchi, L., 2014. Lignin as co-product of second generation bioethanol production from ligno-cellulosic biomass. *Energy Procedia*, 45, pp.52-60.
25. Das, A., Datta, A.K., Bhattacharya, A., Bhattacharyya, A. and Ghose, S., 2010. EMS induced mutagenesis in Poshita and Jawahar 22 of *Withania somnifera (L.) Dunal* (Solanaceae). *Cytologia*, 75(3), pp.305-311.
26. Datta, S.K., 2009. A report on 36 years of practical work on crop improvement through induced mutagenesis. *Induced plant mutations in the genomics era*, pp.253-256.
27. Dave, P.N. and Bhandari, J., *International Journal of Chemical Studies*.
28. De Wild, P.J., Huijgen, W.J.J., Kloekhorst, A., Chowdari, R.K. and Heeres, H.J., 2017. Biobased alkylphenols from lignins via a two-step pyrolysis–Hydrodeoxygenation approach. *Bioresource technology*, 229, pp.160-168.
29. Demirbaş, A., 2001. Relationships between lignin contents and heating values of biomass. *Energy conversion and management*, 42(2), pp.183-188.
30. Devi, M.P., Sahoo, M.R., Kuna, A., Deb, P., Dasgupta, M. and Prakash, N., 2018. Effect of gamma irradiation on nutritional properties and antinutrient contents of *Citrus jambhiri Lush*. *Fruits. Journal of Pharmacognosy and Phytochemistry*, 7(4), pp.2833-2836.
31. EAW (Economic Advisor's Wing, Ministry of Finance). 2013. *Pakistan Economic Survey 2012-13*. Islamabad: Ministry of Finance.
32. Eckstein, D., Künzel, V., Schäfer, L. and Wings, M., 2019. *Global climate risk index 2020*. Bonn: Germanwatch, pp.1-50.

33. Emaga, T.H., Robert, C., Ronkart, S.N., Wathelet, B. and Paquot, M., 2008. Dietary fibre components and pectin chemical features of peels during ripening in banana and plantain varieties. *Bioresource technology*, 99(10), pp.4346-4354.
34. Eshun, J.F., Potting, J. and Leemans, R., 2012. Wood waste minimization in the timber sector of Ghana: a systems approach to reduce environmental impact. *Journal of Cleaner Production*, 26, pp.67-78.
35. Essa, M., S. Babar and Z. Uddin. 2017. Mesquite (*Prosopis juliflora*) plant prevalence in different districts of Balochistan & antibacterial activity of its pods. *Pure Appl. Biol.*, 6(4):1503-1515, December 2017.
36. FAO. 1997b. Asia-Pacific Forestry Sector Outlook Study. (Working Paper Series No. 34). Forestry Policy and Planning Division, Regional Office for Asia and the Pacific, Bangkok.
37. FAO. 2009b. State of the World's Forests 2009. Food and Agriculture Organisation.
38. FAO. 2010. Global Forest Resources Assessment 2010. Country Report Pakistan, Forestry Department FAO- FRA 2010/158, Rome, 2010.
39. FAO. 2016. FAO Soils Portal. <http://fao.org/soils-portal/soilmanagement/management-of-some-problem-10-soils/salt-affected-soils/more-information-salt-affected-soils/en/>.
40. Felker, P. and Bandurski, R.S., 1979. Uses and potential uses of leguminous trees for minimal energy input agriculture. *Economic Botany*, 33, pp.172-184.
41. Fu, X.L., Z.S. Zhang, P. He, X.J. Ou and Q.Y. He. 1999. The effect of gamma ray irradiation on the growth and regeneration of *Cymbidium sinense* rhizome. *Acta Agriculturae Nucleatae Sinica*, 14(6):333-336.
42. Fuwape, J.A., 1989. Gross heat of combustion of Gmelina (*Gmelina arborea* (Roxb)) chemical components. *Biomass*, 19(4), pp.281-287.
43. Galiwango, E., Rahman, N.S.A., Al-Marzouqi, A.H., Abu-Omar, M.M. and Khaleel, A.A., 2018. Klason method: an effective method for isolation of lignin fractions from date palm biomass waste. *Chem. Process Eng. Res*, 57, pp.46-58.
44. Ganesh, B.K., 2011. Effect of mutagen on pollen fertility and other parameter in horse gram [*Macrotyloma uniflorum* (Lam) Verdcourt]. *Bioscience Discovery Journal*, 2(1), pp.146-150.
45. Girija, M. and Dhanavel, D., 2009. Mutagenic effectiveness and efficiency of gamma rays, ethyl methane sulphonate and their combined treatments in cowpea (*Vigna unguiculata* L. Walp). *Global J. Mol. Sci*, 4(2), pp.68-75.
46. Gírio, F.M., Fonseca, C., Carvalheiro, F., Duarte, L.C., Marques, S. and Bogel-Łukasik, R., 2010. Hemicelluloses for fuel ethanol: a review. *Bioresource technology*, 101(13), pp.4775-4800.
47. Girondot, M. 2019. HelpersMG: Tools for analyses in population and evolutionary genetics. R package version 1.8.1.
48. Gnanamurthy, S., Dhanavel, D. and Chidambaram, A.L.A., 2012. Frequency in germination studies of chlorophyll mutants in effectiveness and efficiency using chemical mutagens. *Int. J. Current Life Sci*, 2, pp.23-27.
49. Goel, V.L. and Behl, H.M., 2005. Induced variations, selections and germplasm conservation in selected tree species for afforestation programs on degraded soil sites. *Asian journal of plant sciences*, 4(3), pp.264-270.

50. Gomez, K.A. and Gomez, A.A., 1984. Statistical procedures for agricultural research. John Wiley & Sons.
51. Gonzales, M.A., 2007. Radiosensitivity of three species of ground orchids (*Spathoglottis plicata*, *S. kimbali* var. *angustifolia* and *S. tomentosa*) to acute gamma radiation.
52. Gyimah, L.A., Amoatey, H.M., Boatman, R., Appiah, V. and Odai, B.T., 2020. The impact of gamma irradiation and storage on the physicochemical properties of tomato fruits in Ghana. *Food Quality and Safety*, 4(3), pp.151-157.
53. Hanafy, R.S. and Akladios, S.A., 2018. Physiological and molecular studies on the effect of gamma radiation in fenugreek (*Trigonella foenum-graecum L.*) plants. *Journal of Genetic Engineering and Biotechnology*, 16(2), pp.683-692.
54. Haq, I., Mazumder, P. and Kalamdhad, A.S., 2020. Recent advances in removal of lignin from paper industry wastewater and its industrial applications—A review. *Bioresource Technology*, 312, p.123636.
55. Howard, E.T., 1973. Heat of combustion of various Southern Pine materials. *Wood Science*, 5(3), pp.194-197.
56. Ibrahim, A.M., 2008. Effect of gamma radiation on strawberry genotypes production using tissue culture techniques (Doctoral dissertation, Ph. D. Theses, Fac. Agric., Ain Shams Univ. Cairo, Egypt).
57. Ihsanullah, I., Iqbal, Y. and Khattak, T.N., 2005. Effect of various irradiation doses on some nutrients of Pakistani date. *Journal of Radioanalytical and Nuclear Chemistry*, 266(2), pp.361-366.
58. Irfaq, M. and Nawab, K., 2001. Effect of gamma irradiation on some morphological characteristics of three wheat (*Triticum aestivum L.*) cultivars. *J. Biol. Sci.*, 1(10), pp.935-945.
59. Kalia, C.S., Kharkwal, M.C., Singh, M.P. and Vari, A.K., 2001. Mutagenic effects of environmental industrial chemical agents in inducing cytogenetical changes in wheat. *Indian Journal of Genetics and Plant Breeding*, 61(03), pp.203-208.
60. Kaltschmitt, M. and Streicher, W., 2009. *Energie aus biomasse* (pp. 339-532). Vieweg+Teubner.
61. Katak, R. and Konwer, D., 2001. Fuelwood characteristics of some indigenous woody species of north-east India. *Biomass and Bioenergy*, 20(1), pp.17-23.
62. Kaul, B.L. and Bradu, B.L., 1971. STUDIES ON THE INDUCED MUTATIONS IN MEDICINAL PLANTS. *Planta medica*, 20(05), pp.205-210.
63. Khan, R.A. and Mehmood, R.T., 2003. Potential and prospects of mountain forests. *Mountains of Pakistan: Protection, potential and prospects*, pp.58-72.
64. Khan, S. and Goyal, S., 2009. Improvement of mungbean varieties through induced mutations. *African Journal of Plant Science*, 3(8), pp.174-180.
65. Koyuncu, C. and Yilmaz, R., 2009. The impact of corruption on deforestation: a cross-country evidence. *The Journal of Developing Areas*, pp.213-222.
66. Krauss, A., Krauss, H., Waliszewska, B. and Piatek, J., 2011. Selected environmental aspects of the introduction into the Polish market of exotic wood species on the example of caviuna (*Machaerium scleroxylon Tul.*). *Annals of Agricultural and Environmental Medicine*, 18(2).
67. Kwon, M. and Choe, S., 2005. Brassinosteroid biosynthesis and dwarf mutants. *Journal of Plant Biology*, 48, pp.1-15.

68. Lal, R., 2009. Sequestering carbon in soils of arid ecosystems. *Land Degradation & Development*, 20(4), pp.441-454.
69. Laskar, R.A., Chaudhary, C., Khan, S. and Chandra, A., 2018. Induction of mutagenized tomato populations for investigation on agronomic traits and mutant phenotyping. *Journal of the Saudi society of agricultural sciences*, 17(1), pp.51-60.
70. Liu, C., Wang, J., Huang, T., Wang, F., Yuan, F., Cheng, X., Zhang, Y., Shi, S., Wu, J. and Liu, K., 2010. A missense mutation in the VHYNP motif of a DELLA protein causes a semi-dwarf mutant phenotype in *Brassica napus*. *Theoretical and Applied Genetics*, 121, pp.249-258.
71. Maan, S.S. and Williams, N.D., 1984. An EMS Induced Dominant Allele for Male-Sterility Transferred to Euplasmic Wheat 1. *Crop science*, 24(5), pp.851-852.
72. Mahony, D., 1994. *Trees of Somalia: A field guide for development workers*. Oxfam GB.
73. Maluszynski, K.N., L.V. Zanten and B.S. Ahlowalia. 2000. Officially released mutant varieties, The FAO/IAEA Database. *Mut. Breed. Rev.* 12:1-12.
74. Marcu, D., Cristea, V. and Daraban, L., 2013. Dose-dependent effects of gamma radiation on lettuce (*Lactuca sativa* var. *capitata*) seedlings. *International Journal of Radiation Biology*, 89(3), pp.219-223.
75. Marizka, D., 2010. Uji Ketahanan Tanaman Kedelai (*Glycine max (L.) Merrill*) Hasil Radiasi Sinar Gamma (M2) Pada Cekaman Aluminium Secara In Vitro (Doctoral dissertation, Universitas Sumatera Utara).
76. Mehetre, N.B. and Kshirsagar, R.M. 2022. Effect of Gamma Rays on Germination and Mortality Percentage in M1 And M2 Generation. *Int.J.Curr.Microbiol.App.Sci.* 11(3): 237-240. doi: <https://doi.org/10.20546/ijemas.2022.1103.029>
77. Melki, M. and Sallami, D., 2008. Studies the effects of low dose of gamma rays on the behaviour of chickpea under various conditions. *Pakistan Journal of Biological Sciences: PJBS*, 11(19), pp.2326-2330.
78. Miller, P.D., Vaughn, K.C. and Wilson, K.G., 1984. Ethyl methanesulfonate-induced chloroplast mutagenesis in crops: induction and ultrastructure of mutants. *Journal of Heredity*, 75(2), pp.86-92.
79. Mohammadzai, I.U., Shah, Z., Ihsanullah, I., Khan, H., Khan, H. and Rashid, H., 2010. Effect of gamma irradiation, packaging and storage on the nutrients and shelf life of palm dates. *Journal of Food Processing and Preservation*, 34, pp.622-638.
80. Molin, U. and Teder, A., 2002. Importance of cellulose/hemicellulose-ratio for pulp strength. *Nordic Pulp & Paper Research Journal*, 17(1), pp.14-19a.
81. Moral-Carcedo, J. and Vicéns-Otero, J., 2005. Modelling the non-linear response of Spanish electricity demand to temperature variations. *Energy economics*, 27(3), pp.477-494.
82. Nelkon, M. and Parker, P., 1995. *Advanced level Physics 7th*.
83. Norfadzrin, F., Ahmed, O.H., Shaharudin, S. and Rahman, D.A., 2007. A preliminary study on gamma radiosensitivity of tomato (*Lycopersicon esculentum*) and okra (*Abelmoschus esculentus*). *Int. J. Agric. Res*, 2(7), pp.620-625.
84. Oduor, N. and Githiomi, J.K., 2013. Fuel-wood energy properties of *Prosopis juliflora* and *Prosopis pallida* grown in Baringo District, Kenya.

85. Ontl, T.A., Janowiak, M.K., Swanston, C.W., Daley, J., Handler, S., Cornett, M., Hagenbuch, S., Handrick, C., McCarthy, L. and Patch, N., 2020. Forest management for carbon sequestration and climate adaptation. *Journal of Forestry*, 118(1), pp.86-101.
86. Orozco, R.S., Hernández, P.B., Ramírez, N.F., Morales, G.R., Luna, J.S. and Montoya, A.J.C., 2012. Gamma irradiation induced degradation of orange peels. *Energies*, 5, 3051–3063.
87. Pakistan, Government of. 2005. Supply and Demand of Fuel Wood and Timber for Household and Industrial Sectors and Consumption Pattern of Wood and Wood Products in Pakistan. Ministry of Environment, Government of Pakistan.
88. Pareek, A.K., Garg, S. and Kuma, M., 2015. *Prosopis cineraria*: a gift of nature for pharmacy. *Int J Pharma Sci Res*, 6(6), pp.958-964.
89. Právělie, R., 2016. Drylands extent and environmental issues. A global approach. *Earth-Science Reviews*, 161, pp.259-278.
90. Purente, N., Chen, B., Liu, X., Zhou, Y. and He, M., 2020. Effect of ethyl methanesulfonate on induced morphological variation in M3 generation of *Chrysanthemum indicum var. aromaticum*. *HortScience*, 55(7), pp.1099-1104.
91. Qazi, I.A., 1994. Pakistan: country and forests. *Handbook of Forestry*, 253.
92. R Core Team. 2022. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>
93. Rao, G.M. and Rao, V.M., 1983. Mutagenic efficiency, effectiveness and factor of effectiveness of physical and chemical mutagens in rice. *Cytologia*, 48(3), pp.427-436.
94. Safriel, U.A.Z., Niemeijer, D., Puigdefabregas, J., White, R., Lal, R., Winsolow, M., Ziedler, J., Prince, S., Archer, E., King, C., Shapiro, B., Wessels, K., Nielsen, T.T., Portnov, B., Reshef, I., Thornell, J., Lachman, E. and McNab, D. 2006. Dryland systems. *Ecosystems and Human Well-Being: Current State and Trends*. Island Press, Washington, DC. pp. 625–656.
95. SAKIN, M.A. and SENCAR, Ö., 2002. Makarnal ı k Buğdayda Triticum durum Desf. Gama I şini ve EMS'nin Farklı Dozları n ı n Klorofil Mutasyonları n ı n Oluşumu Üzerine Etkileri. *Journal of Agricultural Sciences*, 8(01), pp.15-21.
96. Sarker, S., Rahman, M.S., Islam, M.R., Hasna, S. and Islam, M.S., 2014. Effect of gamma radiation on morpho-physiological characters of soybean. *Journal of Environmental Science and Natural Resources*, 7(2), pp.25-30.
97. Sasaki, N., Asner, G.P., Pan, Y., Knorr, W., Durst, P.B., Ma, H.O., Abe, I., Lowe, A.J., Koh, L.P. and Putz, F.E., 2016. Sustainable management of tropical forests can reduce carbon emissions and stabilize timber production. *Frontiers in Environmental Science*, 4, p.50.
98. Satpute, R.A. and Fultambkar, R.V., 2012. Effect of mutagenesis on germination, survival and pollen sterility in M1 generation of soybean [*Glycine max (L.) Merrill*]. *International Journal of Current Trends in Science and Technology*.
99. Sazuka, T., Kamiya, N., Nishimura, T., Ohmae, K., Sato, Y., Imamura, K., Nagato, Y., Koshihara, T., Nagamura, Y., Ashikari, M. and Kitano, H., 2009. A rice tryptophan deficient dwarf mutant, *tdd1*, contains a reduced level of indole acetic acid and develops abnormal flowers and organless embryos. *The Plant Journal*, 60(2), pp.227-241.

100. Shah, A.H., Gill, K.H. and Syed, N.I., 2011. Sustainable salinity management for combating desertification in Pakistan. *International Journal of Water Resources and Arid Environments*, 1(5), pp.312-317.
101. Shah, T.M., Mirza, J.I., Haq, M.A. and Atta, B.M., 2008. Radio sensitivity of various chickpea genotypes in M1 generation I-Laboratory studies. *Pak. J. Bot.*, 40(2), pp.649-665.
102. Shea, K.R. and Carlson, L.W. 1984. Increasing productivity of multipurpose tree species: A blueprint for action. *International Union of Forestry Research Organizations (IUFRO) Planning Workshop for Asia*, 16-28 July 1984
103. Shirasawa, K., Hirakawa, H., Nunome, T., Tabata, S. and Isobe, S., 2016. Genome-wide survey of artificial mutations induced by ethyl methanesulfonate and gamma rays in tomato. *Plant biotechnology journal*, 14(1), pp.51-60.
104. Shoukat H.K. 2013. Pakistan's Energy Vision 2030, in *Solutions for Energy Crisis in Pakistan*, ed. Mushir Anwar (Islamabad, Pakistan: Islamabad Policy Research Institute, 2013), 217.
105. Shukla, R. and Datta, S.K., 1993. Mutation studies on early and late varieties of garden chrysanthemum. *Journal of Nuclear Agriculture and Biology*, 22.
106. Sikder, S., Biswas, P., Hazra, P., Akhtar, S., Chattopadhyay, A., Badigannavar, A.M. and D'Souza, S.F., 2013. Induction of mutation in tomato (*Solanum lycopersicum L.*) by gamma irradiation and EMS. *Indian Journal of Genetics and Plant Breeding*, 73(04), pp.392-399.
107. Sikora, P., Chawade, A., Larsson, M., Olsson, J. and Olsson, O., 2011. Mutagenesis as a tool in plant genetics, functional genomics, and breeding. *International journal of plant genomics*, 2011.
108. Song, Q., Li, J., Duan, H., Yu, D. and Wang, Z., 2017. Towards to sustainable energy-efficient city: A case study of Macau. *Renewable and Sustainable Energy Reviews*, 75, pp.504-514.
109. Sri Devi, A. and Mullainathan, L. 2012. The Use of Ethyl Methanesulfonate to Study the Flower Development in (*Capsicum annuum L.*) Mutants. *Botany Research International*, 5(1): 04-09.
110. Stavi, I. 2019. Seeking Environmental Sustainability in Dryland Forestry. *Forests*, 10(9), 737. doi:10.3390/f10090737.
111. Sterling, C. (1976) *Nepal*. *Atlantic Monthly* 23:8(4), 14–25.
112. Stevens, M.A. and Rick, C.M. 1986. Genetics and breeding. In: *The tomato crop* (Atherton JG and Rudlch J, eds). London: Chapman and Hall; 35-109.
113. Sudha Rao, V., Srirangarajan, A.N., Kamat, A.S., Adhikari, H.R. and Nair, P.M., 1994. Studies on extension of shelf-life of rawa by gamma irradiation. *Journal of Food Science and Technology*, 31(4), pp.311-315.
114. Talebi, A.B., Talebi, A.B. and Shahrokhifar, B. 2012. Ethyl Methane Sulphonate (EMS) Induced Mutagenesis in Malaysian Rice (cv. MR219) for Lethal Dose Determination. *American Journal of Plant Sciences* 3: 1661-1665.
115. Tarar, J.L. and Dnyansagar, V.R. 1980. Comparison of ethyl methane sulphonate and radiation-induced meiotic abnormalities in *Turnera ulmifolia* Linn. Var. *angustifolia* Wild. *Cytologia*, 45:221–231.
116. Thurling, N. and Deplttayanan, V. 1992. EMS Induction of early flowering mutants In spring rape (*Brassica napus*) *Plant Breeding* 108:177-184.

117. Timell, T.E. 1959. Isolation of holocellulose from jack pine (*Pinus banksiana*). Pulp and Paper Mag. Can. 60 (1): T26–28.
118. Trossero, M.A. 2002. Wood Energy: The Way Ahead. Unasylva 53, 3–12.
119. Umavathi, S. and Mullainathan, L., 2014. Mutagenic effect of gamma rays and EMS on seed germination, seedling height reduction and survivability of chick pea (*Cicer arietinum L.*) var. Co-4. International Letters of Natural Sciences, 11(1).
120. Van Lierop, P., Lindquist, E., Sathyapala, S., and Franceschini, G. 2015. Global Forest area disturbance from fire, insect pests, diseases, and severe weather events. Forest Ecology and Management, 352, 78–88. doi:10.1016/j.foreco.2015.06.010
121. Wagenführ, R. 2007. Holzatlas, 6th ed. Fachbuchverlag Leipzig im Carl Hanser Verlag, München.
122. Walter, K.J. and Armstrong, K.V., 2014. Benefits, threats and potential of Prosopis in South India. Forests, Trees and Livelihoods, 23(4), pp.232-247.
123. Wang, S. and Han, G.Y., 2011, August. Research on energy crisis and corresponding policies in Heilongjiang province. In 2011 2nd International Conference on Artificial Intelligence, Management Science and Electronic Commerce (AIMSEC) (pp. 544-547). IEEE.
124. Wang, X., Ma, R., Cui, D., Cao, Q., Shan, Z. and Jiao, Z., 2017. Physio-biochemical and molecular mechanism underlying the enhanced heavy metal tolerance in highland barley seedlings pre-treated with low-dose gamma irradiation. Scientific Reports, 7(1), p.14233.
125. Wi, S.G., Chung, B.Y., Kim, J.S., Kim, J.H., Baek, M.H., Lee, J.W. and Kim, Y.S., 2007. Effects of gamma irradiation on morphological changes and biological responses in plants. Micron, 38(6), pp.553-564.
126. Witt, T.W., Ulloa, M., Pelletier, M.G., Mendu, V. and Ritchie, G.L., 2018. Irrigation's effect and applied selection on the fiber quality of Ethyl MethaneSulfonate (EMS) treated upland cotton (*Gossypium hirsutum L.*). Journal of Cotton Research, 1(1), pp.1-11.
127. Xiao, X.O., Lin, W., Li, K., Feng, X., Jin, H. and Zou, H., 2019. Genome-wide analysis of artificial mutations induced by ethyl methanesulfonate in the eggplant (*Solanum melongena L.*). Genes, 10(8), p.595.
128. Yaping, Y., Tiefu, P., Guoquan, L. and Yaokui, X., 1996. Studies on damage effect of EMS to *Coix lacrymajobi L.* Jilin Nong ye da xue xue bao= Acta Agriculturae Universitatis Jilinensis, 18(1), pp.18-20.
129. Yaqoob, U., 2018. Population distribution and water resources in Pakistan.
130. Yousif O.M. 2012. The potential of dehydrated Enteromorpha, *Prosopis cineraria Linn.* (Leguminosae) pods meal and date pits in formulated feed for bluespot grey mullet fry, Valamugil seheli (Forsskal). Journal of Science and Technology 13, 36–45.
131. Youssef, A.A., Aly, M.S. and Hussein, M.S., 2000. Response of geranium (*Pelargonium graveolens L.*) to gamma irradiation and foliar application of Speed Grow. Egyptian Journal of Horticulture, 27(1), pp.41-53.